

Resveratrol as an anticancer nutrient: molecular basis, open questions and promises

Paola Signorelli, Riccardo Ghidoni*

Laboratory of Biochemistry and Molecular Biology, San Paolo University Hospital, School of Medicine, University of Milan, 20142 Milan, Italy

Received 17 September 2004; received in revised form 11 January 2005; accepted 20 January 2005

Abstract

The polyphenol resveratrol is an anticancer nutrient that was shown to inhibit cancer initiation and promotion [Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997;275:218-20]. The absorption, transport and metabolism of resveratrol will be reviewed as well as its actions in multiple pathways involved in the regulation of the cell cycle and the induction of apoptosis. Resveratrol acts as a selective estrogen receptor modulator (SERM) and regulates proteins involved in DNA synthesis and cell cycle, such as p53 and Rb/E2F, cyclins, cyclin-dependent kinases (CDKs) and their inhibitors. Resveratrol affects the activity of transcriptional factors involved in proliferation and stress responses, such as NF- κ B, AP1 and Egr1. Part of these events is mediated by mitogen-activated protein kinases (MAPKs) and tyrosine kinases (e.g., Src) and leads to the modulation of survival and apoptotic factors [e.g., Bcl2 family members, inhibitors of apoptosis (IAPs), ceramide] as well as enzymes involved in carcinogenesis [cyclooxygenases (COXs), nitric oxide synthase (NOS), phase I and II enzymes]. Moreover, resveratrol affects the expression and the activity of cotranscriptional factors such as p300 and sirtuin 1. Thus, resveratrol potential as an anticancer chemopreventive and chemotherapeutic agent and its implication in the prosurvival versus prodeath pathway induction will be discussed. © 2005 Elsevier Inc. All rights reserved.

Keywords: Resveratrol; Cancer; Nutrient

1. Introduction

Food intake modulates metabolism and health, and supplies a variety of bioactive molecules necessary for life. Throughout the ages, medicine has fought against dietary deficiencies. In the 1800s, vitamin deficiencies caused diseases such as scurvy, whereas today, a low intake of phyto-antioxidants, in association with an excess intake of saturated fats in the Western diet, is linked to cardiovascular disease and cancer. During the past 50 years, advances in medical research have led to the development of an extraordinary array of synthetic molecules designed to offer specific cures. However, diseases such as cancer are not easily treated with a single antitumor agent, since they develop from multistep processes and vary according to the genetic and epigenetic characteristics of individuals. Knowledge of the beneficial health effects of natural

compounds present in vegetables, herbs and roots is an instinct for animals and an ancient tradition for humans. Western countries abandoned such traditions in favor of modern food and drug production, but Chinese and Japanese traditional medicines developed this knowledge over the centuries, and accumulated experience and epidemiological observations that represent today our best insight into cancer therapy.

Natural compounds are administered orally and are normally absorbed and metabolized. It is still unknown whether natural compounds are most effective when assumed daily for cancer prevention or when taken at higher doses and in specific combinations aimed at the cure. It is believed that a varied diet is advantageous in that it offers a mix of natural compounds, achieving additive or even synergistic effects.

Natural compounds fit into mechanism-based approach that targets whole pathways and sets of intracellular events rather than a single enzyme, as do many synthetic drugs. This offers a less specific but perhaps more effective strategy for cancer therapy by inducing combinations of

* Corresponding author. Tel.: +39 2 50323250; fax +39 2 50323045.
E-mail address: riccardo.ghidoni@unimi.it (R. Ghidoni).

effects that may counteract the metabolic alterations related to cancer promotion.

This paper explores the latest evidence for the antitumor properties of resveratrol, a polyphenol produced by plants and most known for being responsible for the beneficial health effects of wine.

2. The polyphenol resveratrol

Polyphenols comprise a large class of antioxidants and include flavonoids, anthocyanins, phenolic acids, lignans and stilbenes. These compounds are all derived from phenylalanine and contain an aromatic ring with a reactive hydroxyl group. Within the subclass of stilbenes, resveratrol is the common term for 3,5,4'-hydroxystilbene. Resveratrol exists in both trans and cis isomeric forms (Fig. 1). Since the trans isomer is by far more commonly found in plants and extensively studied, throughout this review, the term resveratrol refers to 3,5,4'-trans-hydroxystilbene.

In plants, polyphenols (including resveratrol) are generally found as 3-O-β-D-glucosides, called piceids (Fig. 1). Other minor conjugated forms of resveratrol contain 1-2 methyl groups (e.g., 3,5-trimethoxy-4'-hydroxystilbene, called pterostilbene), a sulfate group or a fatty acid. Interestingly, substitutions involve a maximum of two hydroxyl groups, and thus conjugated polyphenols maintain their antioxidant properties. A naturally occurring analog of resveratrol that carries four, rather than three hydroxyl groups, is piceatannol (3,4,3',5'-trans-tetrahydroxystilbene).

2.1. Biosynthesis in plants

Resveratrol is produced by a restricted number of plants (about 31 genera). It is not normally present in large amounts, and it is produced in response to stress; in fact, resveratrol belongs to a class of defense molecules called phytoalexins that protect against infection and damage from exposure to ultraviolet (UV) irradiation [1,2]. Resveratrol is toxic to plant pathogens, but some parasites such as fungi overcome this toxicity through the action of membrane proteins (ABC transporters) that transport the compound out of the cellular compartment [3]. Overproduction of stress response molecules in plants triggers a hypersensitivity reaction that can lead to cell death when the stress cannot be counteracted [4].

Resveratrol biosynthesis, catalyzed by stilbene synthase, consists in the repetitive decarboxylative condensation of a *p*-coumaroyl residue from *p*-coumaroyl-CoA with three C₂-units from malonyl-CoA (Fig. 2). Further reactions conjugate native resveratrol to glucosyl or sulfate residues at the 3-position of the biphenolic ring.

Resveratrol is susceptible to oxidative degradation, while the glycosylated piceid form is resistant. Glycosylated resveratrol maintains its biological activity, is more stable and soluble, and therefore is more readily absorbed by the human intestine [5].

2.2. Natural sources

Resveratrol and the analogs piceatannol and pterostilbene are found in several edible natural products such as grapes (*Vitis* spp.), peanuts (*Arachis* spp.) [6], berries (blueberries, cranberries and lingonberries, all *Vaccinium* spp.) [7] and rhubarb (*Rheum* spp.) [8]. Resveratrol was first detected in the dried roots of *Polygonum cuspidatum* (Itadori tea), traditionally used in Chinese and Japanese medicines as an antiinflammatory agent [1,9]. Moreover, its occurrence in the plant kingdom is widespread, being present in wild nonedible berries of *Vaccinium* spp. [7] and in *Eucalyptus* spp. [10], *Yucca schidigera* [11], *Dracaena loureiri* [12], *Cassia* spp. [13], mulberry (*Morus* spp., *Maclura pomifera*, *Nothofagus fusca* spp.), and red sandalwood (*Pterocarpus* spp., a major source of pterostilbene) [7]. Extracts from roots, heartwood, bark and leaves of most of these plants are commonly used in traditional oriental medicine.

The content of resveratrol in different sources varies widely, depending on factors such as cultivar, climate, fungi infections, UV exposure and wine-making procedures.

2.3. Biological activities of resveratrol: from plants to humans

Many classes of phytochemicals exert antioxidant activities as well as other beneficial effects, for example, on the inflammatory responses, on cellular enzymatic detoxification systems and on proliferative and apoptotic factors. The human diet contains a mixture of plant-derived polyphenols such as genistein, quercetin, epigallocatechin (from soybeans and green tea leaves) and resveratrol, mainly present in grape skin and peanuts. The biological activities of these substances have been demonstrated in humans [14,15]. In the early 1970s, epidemiological studies revealed an inverse correlation between red wine consumption and cardiovascular diseases in France [16]. The “French paradox” was resolved with the identification of antioxidants such as resveratrol in wine. Subsequently, a large number of studies identified multiple beneficial effects of this molecule in humans [16]. In plasma, resveratrol was associated with lipoproteins [17] and it was shown in vitro to inhibit low-density lipoprotein (LDL) oxidation [18,19]. In vitro studies documented that resveratrol inhibited platelet aggregation [20] and polymorphonuclear cell activation (production of reactive oxygen species) [21]. In human endothelial cells, resveratrol induced vasorelaxation [22] and impaired migration and tube formation, thus reducing thrombogenic potential, by inhibiting expression of adhesion molecules [21,23,24].

In the last decades, phenolic acids (e.g., gallic acids, caffeic acids, tannic acids, curcumin), flavonoids (genistein, daidzein, quercetin, myricetin, kaempferol) and other polyphenols (epigallocatechins) were shown to induce apoptosis in cancer cells [14], and a few clinical trials using these natural products have been carried out [25,26]. In 1997, Jang et al. [13] reported that resveratrol exerts antitumor

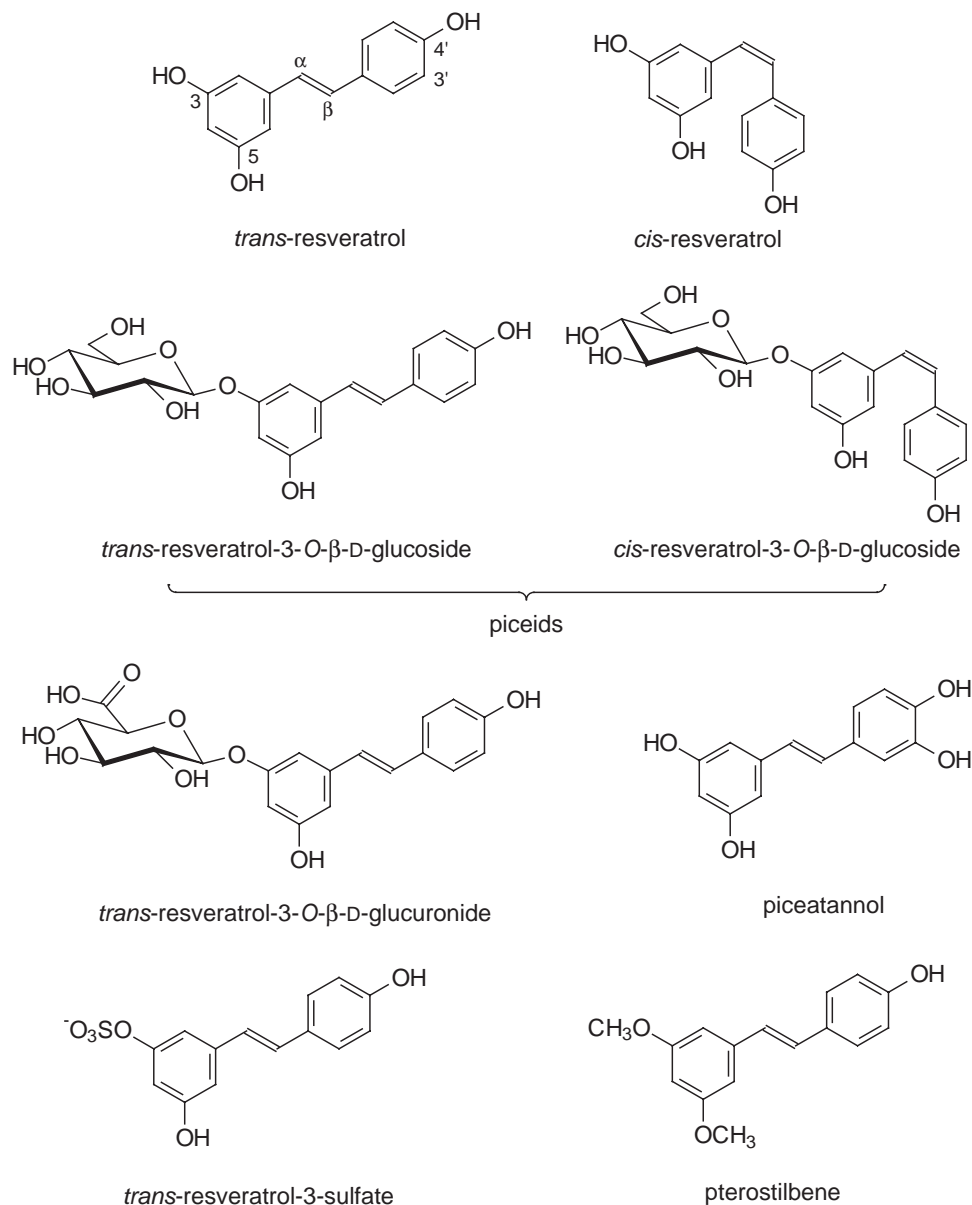


Fig. 1. Structural formulas of resveratrol and its most common conjugates and analogs.

properties by reducing tumor mass in rats. These authors demonstrated that resveratrol is effective in blocking *in vivo* the three stages of carcinogenesis: initiation, promotion and progression. Since this pioneer work was published, many studies employing human cancer cell lines have confirmed this observation and have sustained that resveratrol is a chemopreventive agent [27]. Less convincing and still to be established is its chemotherapeutic potential.

3. Resveratrol in the diet: absorption, transport, metabolism and excretion

In order to understand the potential of specific and beneficial properties of dietary compounds, it is useful to study their absorption after oral intake, transport in body

fluids, cellular metabolism and, ultimately, excretion. The absorption and transport of resveratrol have been studied in several models: isolated rat intestine [28,29], rats and mice after oral administration [30–34], human colon carcinoma Caco-2 cell line [35], human hepatocytes [36] and healthy human subjects [31,37]. The complex pathways for resveratrol absorption, transport, metabolism and excretion are summarized in Fig. 3.

3.1. Absorption

Experiments with isolated rat intestine perfused *in vitro* with resveratrol-containing buffer showed that jejunum and, to a lesser extent, ileum are involved in the absorption of resveratrol [29]. Most of the resveratrol transported to the basolateral side was in a metabolized form mainly as

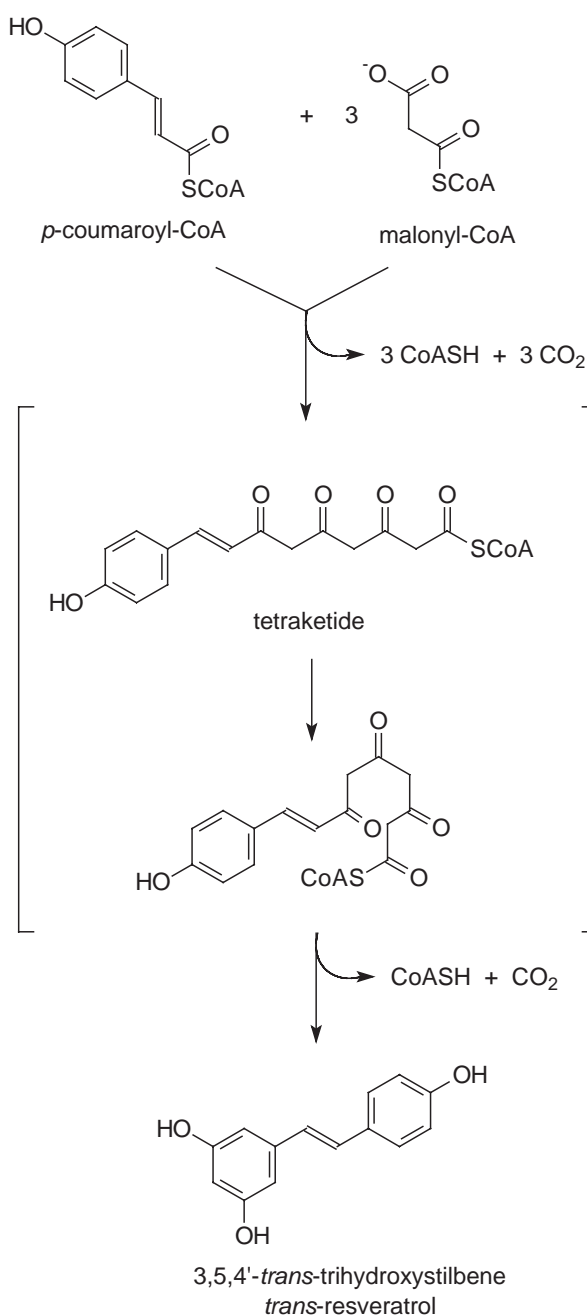


Fig. 2. Biosynthetic pathway of resveratrol.

glucuronide and also as sulfate conjugates. However, the total amount of resveratrol and its conjugates crossing the intestinal epithelium was only 6% of the total perfused amount [29]. The results of this *in vitro* study contrast with the higher intake observed *in vivo*, in both animals and humans; for example, in mice fed orally with resveratrol, up to 75% of the ingested content was absorbed and the remainder was eliminated in the feces [31].

In vitro studies showed that resveratrol is metabolized also by human intestinal cells. The human colonic adenocarcinoma cell line Caco-2, treated *in vitro* with resveratrol, exhibited initially a dose-dependent increasing

rate of apical to basolateral transport, until this reached a plateau with a maximal transport that is not concentration dependent [35]. Reverse transport may also occur when cells are incubated with resveratrol on the basolateral side. Resveratrol is released from enterocytes in a conjugated soluble form. In this study on human cells, resveratrol was released mainly as sulfatide conjugate and only in minimal amount as glucuronide conjugate [35], whereas in rats, the majority of the resveratrol metabolism involves glucuronidation [29,31].

Resveratrol is present in dietary products as *cis*- and *trans*-resveratrol but mainly in glycosylated forms named piceids (3-*O*- β -D-glucosides). Plants and pathogens as well as the human digestive tract contain enzymes that oxidize polyphenols. Glycosylation inhibits enzymatic oxidation of resveratrol, thereby preserving its biological activity and increasing its stability and bioavailability [5,38]. Since intestinal cells are only able to absorb aglycone resveratrol, the absorption process requires glycosidases [39]. Therefore, the relative amounts of aglycone and glycosylated resveratrol in foods may modulate the absorption rate. Different activities and expression levels of intestinal glycosidases may explain the differences in resveratrol uptake rates observed in humans and rats.

3.2. Transport to tissues

Conversion of resveratrol into hydrophilic conjugates may facilitate its entry in the blood stream, its diffusion throughout the body and, most importantly, its excretion.

In vivo studies in animals showed that after the oral administration of resveratrol, both aglycone and conjugated forms appeared in plasma. With time, plasma concentration diminished until a secondary peak appeared. This secondary peak was due to the recirculation of resveratrol after release from bile. The liver and gallbladder filtered resveratrol and its metabolites from the circulation and transported them back again into the intestine through the bile for a delayed absorption [30,33]. Shortly after oral intake, resveratrol was found in colon, whereas its tissue distribution required a few hours. In liver, resveratrol accumulated up to a concentration comparable to that which exerts biological effects in *in vitro* assays (micromolar range) [34].

The uptake and metabolism of resveratrol by human liver have been studied in *in vitro* models. Human hepatocytes exhibited an initially increasing rate of uptake (minutes), then the rate remained stable (hours) [36]. At low concentrations, hepatocyte uptake was mediated by temperature-sensitive active transporters (half saturation at 30 μ M). At higher concentrations, the molecule diffused into cells in a nonsaturable but concentration-dependent process [36].

3.3. Metabolism and excretion

In animal model, renal excretion of resveratrol started within hours after intake and increased throughout the next 12–24 h [34]. The presence in urine of little native (aglycone) resveratrol but high amounts of its conjugates

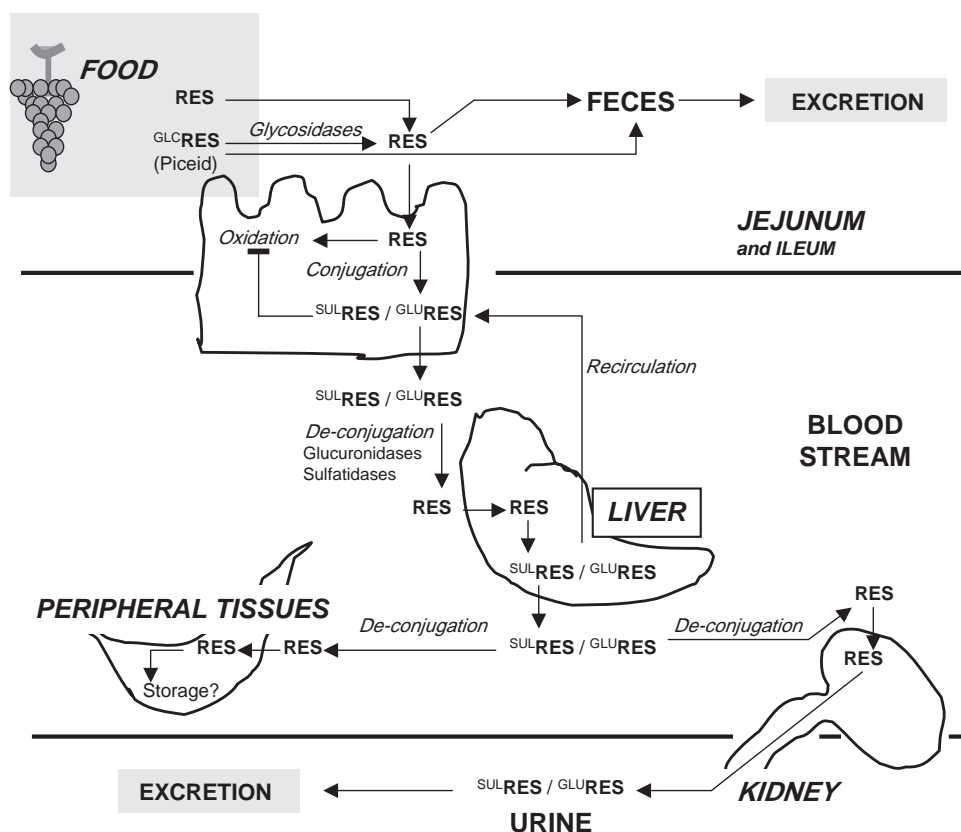


Fig. 3. Pathways of resveratrol absorption, transport, metabolism and excretion. ^{GLC}RES, resveratrol-3-*O*- β -glucoside (piceid); ^{SUL}RES, resveratrol-3-sulfate; ^{GLU}RES, resveratrol-3-*O*- β -glucuronide.

indicates that metabolism of the compound is essential for excretion [33]. In kidney, resveratrol was present mainly in its native form, whereas in urine, the majority of the compound was present in its conjugated form [34].

In human, the excretion time depended on the concentration of resveratrol (aglycone and conjugates) present in plasma, but there was no direct correlation between excreted and introduced amount. This observation from Meng et al. [40] suggests that while small amounts of resveratrol are rapidly metabolized and eliminated, retention and accumulation of the compound in tissues occur only over a certain dose of intake, thus becoming potentially available for cellular uptake and intracellular signaling.

Conjugation with glycidic or sulfatidic groups is probably aimed at promoting excretion, since conjugates are not intracellularly active [41]. Flavonoids such as quercetin inhibit the glucuronidation of resveratrol and therefore may increase its bioavailability [42]. This observation may partly explain why low concentrations of dietary compounds have synergistic effects [43].

Many questions regarding resveratrol metabolism and excretion remain unanswered. It is still to be clarified if resveratrol conjugates are transported to tissues or if they are only targeted for excretion. Moreover, since liver uptake has only been studied *in vitro* using the native aglycone form of

resveratrol, it is not known whether hepatocytes also absorb the conjugated form of resveratrol from the blood stream or if these conjugates are targeted for excretion.

4. Antitumoral activity of resveratrol

Antitumor agents impair procarcinogen metabolic activation and interaction with cellular targets (DNA, proteins); they inhibit cancer development by blocking tumor cell transformation and proliferation and by inducing tumor cell death [14]. Therefore, chemoprevention and chemotherapy can be obtained by acting at multiple levels and by impairing combinatorial effects responsible for mutagenesis and mitogenesis. Among food-derived molecules that have been screened for the ability to inhibit or reverse such cellular processes, resveratrol is particularly interesting because it affects a broad range of intracellular mediators involved in the initiation, promotion and progression of cancer [13,15,27]. Since Jang et al. showed *in vivo* the antitumor potential of resveratrol, many studies revealed a variety of resveratrol intracellular targets whose modulation give rise to overlapping responses that lead to growth arrest and death. Anyway, the efficacy of this molecule is still debated because of the multiplicity of affected targets and contradictory effects related to dose and time of treatment

and to cellular phenotype. The specific cell molecular setting determines the response to resveratrol treatment. In this review, the action of resveratrol on tumor genesis and growth will be examined and discussed.

4.1. Resveratrol: a phytoestrogen with agonistic and antagonistic hormonal activities

Loss of estrogen and androgen production in aging leads to deregulated functioning of tissues and organs. Moreover, improperly balanced hormonal stimulation may favor cell proliferation over differentiation and senescence, and may increase the risk of developing cancer. Consequently, hormone-dependent tumors (breast and prostate, but also others such as colon and lung) may be prevented by the daily intake of appropriate amounts of selective estrogen receptor modulators (SERMs). These compounds exhibit various degrees of estrogen agonism or antagonism, depending on the cell type, expression of genes targeted by estrogen receptors (ERs) and other related intracellular responses [44].

Several polyphenols structurally resemble estrogens. Phytoestrogens such as the flavonoid quercetin and the isoflavone genistein behave as both agonists and antagonists of estrogens receptors, giving rise to opposing responses according to concentration, competition and ER expression [45]. Breast cancer incidence among women in Western countries is sixfold higher than that among women in Asia who consume daily soy products rich in phytoestrogen, suggesting that these latter may act as chemopreventing agents [46].

The polyphenol resveratrol can be considered as a dietary phytoestrogen with powerful beneficial effects on both ER-expressing and nonexpressing human tumors. The chemical structure of resveratrol is similar to that of the synthetic estrogen diethylstilbestrol (4,4'-dihydroxy-trans-diethylstilbene). Resveratrol belongs to the type I class of estrogens [47]. It binds ERs in the low micromolar range with an affinity lower than that of estradiol; therefore, it behaves as a weak competitor. Despite the lower binding affinity, resveratrol may act as a superagonist in activating hormone receptor-mediated gene transcription [47,48]. The superagonistic induction of gene expression is related to the promoter context and varies according to cell type [49].

Aside from superagonism, resveratrol also exerts an antiestrogen action by triggering parallel pathways that inhibit estrogen-induced cellular outcomes, such as proliferation, tumoral transformation and progression [13,50,51]. In the presence of estrogen, resveratrol exerted a mixed agonistic–antagonistic action in ER-positive breast cancer cells [52]. Resveratrol binding to ER β suppressed the expression of the α form of ER [53] and regulated androgen receptor signaling by repressing receptor coactivators and downstream gene transcription [54,55]. Kuwajerwala et al. [56] reported that in androgen-sensitive prostate cancer cells, resveratrol induced a proliferative effect at low dose

(5 μ M) but an apoptotic effect at 15 μ M or higher dose, whereas in androgen-independent cells, resveratrol did not increase DNA synthesis at any concentration. It has been proposed that resveratrol inhibition of focal adhesion kinase (FAK) and protein kinase B (PKB/Akt) is responsible for inducing apoptosis in ER-positive breast cancer cells [57]. In this study, apoptosis occurred only in ER-positive cells but it did not involve receptor-mediated gene transcription. Steroids (both estrogens and androgens) and their receptor complexes modulate mitogen-activated protein kinases (MAPKs) [by activating ERK, down-regulating Jun N-terminal kinases (JNK)] and thus downstream transcriptional events [58]. Mitogen-activated protein kinases have been shown to mediate also upstream events induced by resveratrol, leading to downstream regulation of transcriptional factors [59–61]. In breast cancer cells, 17 β -estradiol reversed resveratrol-induced apoptosis even if estrogens and resveratrol acted similarly in stimulating MAPKs (specifically ERKs), either when administered alone or in combination (additive effect) [62].

This contrasting evidence may be explained by considering resveratrol ability to alter nongenotropic activities of steroid receptor complexes. Activated ER may rapidly stimulate the activity of G proteins and protein kinases, independently of gene transcriptional events [63].

Resveratrol may therefore be considered as a natural SERM [49], although the balance between prosurvival genotropic and opposing nongenotropic activities is not clearly predictable due to the role of a broad array of intervening factors. At the low doses provided by dietary intake, resveratrol may act as a weak estrogen competitor, according to the receptor expression and hormonal status of tissues; it counteracts the proliferative effects of hormones and provides a balancing antitumoral activity. Tissue-specific expression of α and β ERs, cofactors regulating DNA binding and different gene promoters, all modulate the cellular response to resveratrol. In the absence of endogenous hormones and according to cellular specificity, the superagonistic activity of resveratrol may act in an opposite manner and prevent tissue senescence and apoptosis. When stress signals overcome proliferative signals, or when these latter are missing (absence of hormones), the polyphenol-induced pathway may switch to apoptosis (Fig. 4).

Aggressive breast and prostate tumors often lose ER expression and become estrogen independent. However, resveratrol is also able to affect ER negative cells. Besides binding hormone receptors, resveratrol is able to trigger intracellular events that affect different metabolic pathways. Indeed, resveratrol exhibits antitumoral properties in a variety of cells that do not express steroid hormone receptors [51,52,64], as well as in cells treated with ER blockers (e.g., tamoxifen) [65]. These observations indicate that resveratrol triggers multiple pathways that may or may not involve ER activation. In this sense, resveratrol may be considered as both a chemopreventive and a chemotherapeutic agent.

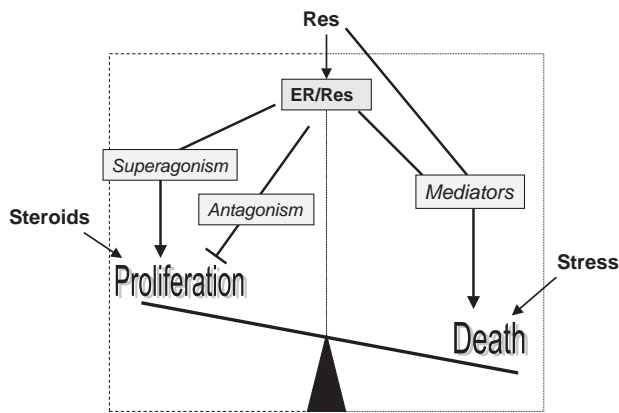


Fig. 4. Pathways in which resveratrol acts as a SERM.

4.2. Resveratrol: a modulator of phase I and phase II enzymes

Cancer initiation occurs as a consequence of multiple events. The combined effects of stimulating factors (e.g., hormones, cytokines), stress mediators (inflammatory oxygen radicals) and exogenous aggressions (viruses, radiation and xenobiotic compounds) can affect the control of cellular proliferation and lead to the tumoral transformation of tissues.

Xenobiotics (carcinogens and drugs) are often lipophilic substances that easily enter cells. In order to protect from their toxicity, cells render them more hydrophilic and excretable through a process called biotransformation. This process involves oxidation by phase I enzymes and conjugation with polar groups by phase II enzymes. However, this process may lead to the formation of highly reactive oxygen radicals and other molecules that easily interact with DNA and thus procarcinogens become converted into carcinogens.

The oxidative phase involves membrane-bound enzymes (e.g., cytochrome P450 monooxygenase, CYP) that use molecular oxygen to introduce an oxygen atom into substrates [66,67]. The second phase is carried out by transferases that add hydrophilic groups or change the redox state of the molecule (e.g., glutathione-S-transferase, UDP-glucuronosyltransferase, sulfotransferase, NAD(P)H:quinone oxidoreductase). Chemopreventive strategies include the inhibition of phase I enzymes responsible for activating xenobiotics and the induction of phase II enzymes that conjugate these activated compounds to endogenous ligands (e.g., glutathione).

Resveratrol, being an exogenous lipophilic compound, can cross plasma membrane, be subjected to cellular metabolism and it possibly interacts with phase I enzymes. Resveratrol inhibited human recombinant CYP P450 in vitro [41]. Moreover, it inhibited CYP450 activity from mouse or human liver microsomes [68,69]. Jang et al. [13] found that resveratrol reduced the insurgence of preneoplastic lesions in mouse mammary gland cultures and decreased the incidence of tumor formation in mice treated with 7,12-dimethylbenz[*a*]anthracene (DMBA) used as

tumor initiator, in combination with phorbol esters used as tumor promoter. Since DMBA requires bioactivation by phase I enzymes CYP1A1, CYP1A2 and CYP1B1 [70], the antitumoral activity of resveratrol in vivo includes prevention of the initiation phase of carcinogenesis by inhibiting phase I enzymes. Resveratrol was found to be able to discriminate among CYP isoenzymes: it inhibited CYP1A1 and CYP1B1 activities directly while it inhibited CYP1A2 indirectly. In this study, it is suggested that indirect CYP inhibition is due to a compound derived from resveratrol metabolism in the presence of NADPH [71].

Besides inhibiting CYP activity, resveratrol also acted at the transcriptional level by blocking the activation of CYP1A1 promoter and gene transcription in human hepatoma cells [69]. In addition, resveratrol impaired the carcinogenic effect of aryl-hydrocarbons. Aryl-hydrocarbons are carcinogens that act via nuclear receptors to promote CYP transcription for the enzymatic conversion of xenobiotics in carcinogen elements. Resveratrol, without binding the receptors, impaired their interaction with the promoter region of the CYP1A1 gene [69].

Resveratrol was further shown to induce phase II enzymes such as UDP-glucuronyltransferase and NAD(P)H:quinone oxidoreductase in mouse epidermis [72].

These data strengthen the hypothesis that resveratrol may be used in cancer prevention. The effects of resveratrol on phase I and II enzymes are summarized in Fig. 5.

4.3. Resveratrol: modulator of nitric oxide synthase

Nitric oxide synthase (NOS) is a heme-containing monooxygenase with a reductase domain and an oxygenase domain. Constitutive isoforms (endothelial, neuronal and mitochondrial NOS) provide low intracellular concentrations of the short-lived free radical nitric oxide (NO). An inducible form of NOS (iNOS), activated at the transcriptional level during inflammation, provides high concentrations of NO

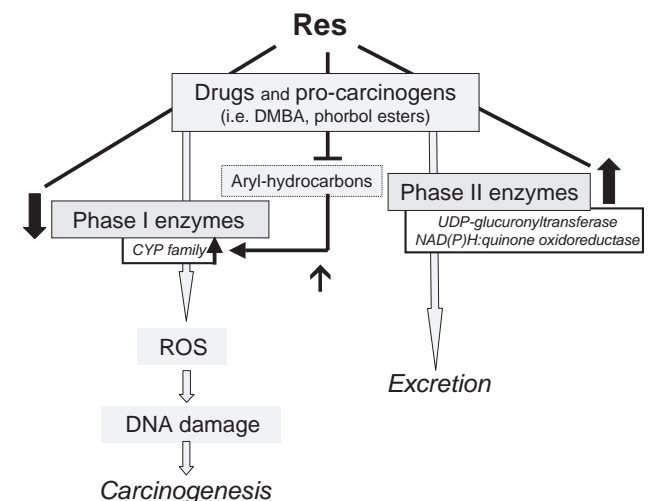


Fig. 5. Pathways in which resveratrol modulates phase I and phase II enzymes.

[73]. NO is known for its vasodilatory properties [74], its inhibition of platelet adhesion and aggregation [75], and of adhesion molecule expression [76], and its suppression of cell growth and migration [77]. It is thus apparent that NO and resveratrol share common targets.

High levels of endogenously produced NO may either induce apoptosis (by generating toxic reactive intermediates) [78,79] or inhibit it [80,81], depending on the intracellular redox state. The promoter region of iNOS is controlled by transcription factors such as NF- κ B, AP1 (Jun/Fos), CREB and STATs. The expression of iNOS is often associated with the induction of tumoral markers such as cyclooxygenase 2 (COX2), vascular endothelial adhesion molecule 1 (VCAM1) and intercellular adhesion molecule 1 (ICAM1) [82]. Finally, iNOS is up-regulated in cancer cells in relation to tumor progression [73,83].

Resveratrol may favor antitumoral activity by impairing the process of angiogenesis. The compound was shown to inhibit endothelial cell migration and tube formation and to block oxygen radical formation and the related Src-mediated expression of vascular endothelial cadherins [23]. Resveratrol treatment in the micromolar range induced vascular smooth muscle cell relaxation by promoting NO release, and this was reversed by NOS inhibitors. The action was indirect since resveratrol had no effect on NOS activity in rat aortic homogenates measured *in vitro*. Moreover, metabolic inactivation of NO by NADH/NADPH oxidase was inhibited by resveratrol, thus potentiating the effect [84]. Moreover, resveratrol induced the expression of the endothelial isoform of NOS (eNOS), in association with the activation of p53 and p21 and cell cycle arrest in S/G2 phase [85].

Conversely, resveratrol was shown to have opposite effects in cancer cells. It inhibited NO production and iNOS expression while it also induced apoptosis in human B-cell lines derived from patients with chronic B-cell malignancies or lymphocytic leukemia [83]. Similarly, in lipopolysaccharide (LPS)-stimulated macrophages, resveratrol reduced iNOS mRNA transcription and cytosolic protein levels, possibly by blocking phosphorylation of I- κ B and activation of NF- κ B [86].

Although there is conflicting evidence regarding the effect of resveratrol on NO formation, it appears to have antitumoral action via the inhibition of angiogenesis or the induction of cell cycle arrest and apoptosis in cancer cells.

4.4. Resveratrol: a modulator of COX

Resveratrol was first discovered in roots for its ability to inhibit the activity of COXs, enzymes that catalyze the first committed steps of prostaglandin (PG) biosynthesis. Prostaglandins are known stimulators of cell proliferation and angiogenesis, and suppressors of immune surveillance [87]. Cyclooxygenases also possess a hydroperoxidase activity that converts PGG₂ to PGH₂; this activity, which generates tyrosyl radicals, may be involved in the bioactivation of promutagens, as are phase I enzymes [88].

Resveratrol noncompetitively inhibited both the COX and hydroperoxidase activities of COX1 *in vitro* [13,89]. This dual effect of resveratrol is unique, since classic nonsteroidal antiinflammatory drugs (NSAIDs) only affect the COX activity [90,91].

Besides constitutive COX1, which is ubiquitously expressed and produces PGs involved in tissue homeostasis, an inducible form of COX2 is expressed during inflammation and neoplasia; COX2 is induced by proinflammatory and mitogenic stimuli [92]. Since overexpression of COX2 inhibits apoptosis and promotes metastasis of tumor cells [93,94], targeting COX2 expression could be a promising chemotherapy strategy.

Resveratrol inhibited human recombinant COX2 in *in vitro* assays [95]. Resveratrol discriminated between the two COX isoforms, being poor inhibitor of COX2 hydroperoxidase activity [13,89]. The two COX isoforms contribute to the formation of thromboxane A₂ (TXA₂), and PGI₂ from PGH₂ [96] and resveratrol, with its selective action, may produce different effects according to tissue expression and functions of the isoforms.

Despite *in vitro* data on enzyme inhibition, the most relevant mechanism occurring *in vivo* may likely be a mediated one. Down-regulating COX2 transcription [97], resveratrol was proposed in breast cancer cells to act by inhibiting the activation of the transcriptional factor NF- κ B, upstream of COX2 expression [98]. In addition, in a similar cell model, micromolar concentrations of resveratrol blocked phorbol-ester-mediated translocation of protein kinase C (PKC) to the membrane and activation of the COX2 promoter [95]. This study showed that resveratrol inhibited the activation of a cyclic-AMP-responsive element (CRE) that controls COX2 expression, and that mutation of this region abrogated the effects of resveratrol.

The currently available evidence indicates that resveratrol behaves as a tumor-promotion antagonist, by triggering intracellular pathways that oppose the unbalanced expression of COX2.

4.5. Resveratrol-induced growth arrest and apoptosis in cancer cells

Cancer cells escape from cell cycle control and from G₀/G₁ terminal differentiation. Resveratrol was found in a variety of cell models to arrest proliferation, mostly in an irreversible way, leading to apoptosis. This effect has been traced to resveratrol ability to modulate the activity of many key mediators of cell cycle and survival.

4.5.1. Cell cycle-regulating proteins

Several protein kinases [cyclin-dependent kinases (CDKs)], their activators (cyclins) and their associated inhibitors form a network of complexes driving cells in and out of the cell cycle phases; their action during the cell cycle is switched off by sequential degradation [99].

A number of studies reported that a variety of different human cancer cell lines, treated with resveratrol at micro-

molar concentrations for 12–24 h, arrested their proliferative cycle in the G1/S boundary [100], in the S phase [98,101–109] or, less frequently, in the G2/M phase [110,111]. One subject of debate is whether resveratrol-induced cell cycle arrest is reversible or is the first step of an irreversible apoptotic program. A number of studies found that the resveratrol-induced cell cycle block did not cause apoptosis [101,103,106,112], was reversible [101,102,112,113] and was associated with cell differentiation in a variety of cancer cells [101,112,114,115].

Cell sensitivity to apoptotic agents may change throughout cell cycle phases [116]; therefore, resveratrol ability to arrest cancer cells in S phase, which is the most vulnerable, may strongly increase its chemotherapeutic potential [109].

Moreover, many authors reported that the induction of cell cycle arrest by resveratrol was followed by apoptotic cell death [56,84,100,104,106–108,111,117,118].

A number of protein targets of resveratrol have been identified. Among these, pivotal roles in cell cycle progression, either in normal or in stressed conditions, are exerted by p53 and by the retinoblastoma gene product (Rb). The p53 oncosuppressor is a DNA-binding protein that activates transcription of genes that induce cell cycle arrest. p53 is present at low levels in normally proliferating cells, and it accumulates, as a result of increased stability due to acetylation and phosphorylation, in response to stress and in senescence [119–122]. p53 arrests the cell cycle in G1 by activating the cyclin inhibitor p21 [123,124]. The outcome of p53-arrested cells is apoptosis mediated by mitochondrial Bcl family proteins, resulting from the increased expression of proapoptotic factors and the activation of caspases [125].

In a variety of cellular models, resveratrol strongly up-regulated p53 and p21, imposing a checkpoint at G1/S transition [84,100,103,106], although the compound was also reported to act independently from p53 cellular status [109]. This altered equilibrium led to the modulation of CDKs and cyclins, both at transcriptional and posttranscriptional levels, resulting in cell cycle arrest in a specific phase [55,56,84,100,102,106,126]. Narayanan et al. demonstrated that resveratrol activated transcription of a whole set of p53-responsive genes (e.g., p21, p300/CBP, Apaf1 and BAK) related to cell cycle arrest and apoptosis, while it down-regulated tumor-associated antigens (e.g., PSA), NF- κ B/p65 and Bcl2 [127]. Resveratrol not only increased p53 cellular content but also induced its posttranslational modification (phosphorylation and acetylation) required for regulating gene transcription (such as p21 induction) [62]. Interesting observations have been reported on endothelial cell proliferation and angiogenesis, a process that plays an important role in tumor growth. In vascular endothelial cells, resveratrol activated p53 via serine phosphorylation but without increasing total concentration of p53 or p21 [102]; thus, resveratrol induced a block in DNA synthesis that was not followed by apoptosis. These authors hypothesized that this partial activation of p53 reversibly blocked

proliferation because washout of resveratrol restored normal cell cycle progression. In a similar endothelial cell model, other authors observed that resveratrol activated p53 and p21 and induced cell cycle arrest, but this arrest led to apoptosis only in proliferating serum-stimulated cells and not in quiescent cells [103]. This observation is in agreement with other studies reporting that resveratrol was differently toxic to tongue squamous carcinoma than gingival fibroblasts [128], and that it was toxic to leukemia cells but not to normal hematopoietic progenitors [129] or peripheral blood lymphocytes [130].

The regulation of p53 by resveratrol has been proposed to occur via activation of MAPKs (specifically ERKs and p38) [131]. Mitogen-activated protein kinases family members are implicated in cellular proliferation and also in apoptosis [132]. Resveratrol activated p38 and inhibited JNK in stimulated human cervical cancer cells [60], and it activated ERK1/2 in human breast cancer cells [62], human erythroleukemic cells [101] and human melanoma [133].

Moreover, p53 activity was shown to be regulated by resveratrol via modulation of p300 expression [54,127]. p300 is a transcription coactivator and member of the CREB-binding proteins family; it has acetyl-transferase activity that structurally alters many transcription factors, thereby exerting an antitumor role [134]. Targets of p300 include transcriptional factors known to be regulated by resveratrol, such as NF- κ B, p53 [135] and Egr1 [136]. It is possible that p300 and p53 belong to an apoptotic loop of gene regulation that mediates resveratrol-induced apoptosis.

The retinoblastoma gene (Rb) product plays an important “gate keeper” role in the G1/S transition in the normal homeostatic situation. In the hypophosphorylated active form, Rb associates with E2F family of transcriptional factors, repressing gene transcription. By the end of G1 phase, hypophosphorylated Rb becomes hyperphosphorylated and inactivated by CDK/cyclin complexes [137], releasing E2F proteins. E2F complexes stimulate the transcription of genes that promote entry into S phase (including survivin, another target of resveratrol, as will be discussed later). Resveratrol action on Rb phosphorylation is controversial. In rat vascular smooth endothelial cells, high resveratrol concentration (100 μ M) increased Rb phosphorylation and favored the transition into phase S but reversibly arrested the cell cycle in this phase, possibly via p53, although cellular levels of cyclin inhibitors p21 and p27 decreased [102]. Conversely, prolonged treatment (24–48 h) of human epidermoid carcinoma cells with low concentrations of resveratrol (10 μ M) decreased the phosphorylation of Rb and the expression of E2F proteins and their complex-forming partners DP proteins, by up-regulation of p21 and the block of related CDK/cyclin complex, leading to cell cycle arrest in G0/G1 [118]. Interestingly it has been recently demonstrated that accumulation of free E2F may favor apoptosis by stabilizing and thus increasing p53 [138].

Another mechanism involved in resveratrol block of the cell cycle is related to its inhibition of DNA synthesis,

thereby impairing the normal course of the S phase. In particular, resveratrol inhibited ribonucleotide reductase activity in leukemia cells [139] and inhibited DNA polymerase activity in vitro [140].

Indeed, resveratrol ability to potentiate the effectiveness of apoptotic or cytostatic drugs requires pre- or cotreatment, whereas the addition of resveratrol after stressing stimulus is ineffective [109,141]. In vitro experiments demonstrated that resveratrol inhibited DNA polymerase, protein kinase D (PKD), cPKCs and COX2, although the concentration of the compound used in these in vitro assays was much higher than the tissual/cellular concentration available with dietary assumption or pharmacological treatment of resveratrol.

4.5.2. Cell survival-related proteins

Resistance to apoptosis may depend on intracellular levels of prosurvival and proapoptotic factors such as members of the Bcl2 family and the inhibitors of apoptosis (IAPs) proteins family.

The Bcl2 family of proteins includes pro- and anti-apoptotic factors. Proapoptotic proteins activate caspases, and this event is prevented by heterodimerization of proapoptotic with antiapoptotic proteins. Heterodimer formation is regulated by phosphorylation [142]. Overexpression of Bcl2 family proteins impairs resveratrol-induced apoptosis in T-acute lymphoblastic leukemia [143] and significantly attenuated resveratrol-induced apoptosis in monocytic leukemia [144], by impairing alterations in mitochondrial membrane permeability, production of radical oxygen species (ROS) [143], cytochrome C release and caspases activation [144].

Recently, it has been shown that natural polyphenols (e.g., catechins, epigallocatechins, theaflavins) bind Bcl2 and BclX1; the binding impaired the ability of these proteins to balance proapoptotic family members and thus it induced apoptosis [145]. Given structural similarities between these polyphenols and resveratrol, it will be interesting to study whether resveratrol is also able to interact with and sequester antiapoptotic factors. In colon carcinoma cancer cells, resveratrol induced mitochondrial translocation of proapoptotic Bcl2 family members (e.g., Bax, Bak) and initiated an apoptotic cascade [146]. Moreover, resveratrol treatment down-regulated the expression of antiapoptotic and increased that of proapoptotic Bcl2 members [127,146].

Survivin, a member of the IAP family, directly inhibits apoptosis, and its expression is frequently high in cancer cells and correlates with resistance to chemotherapy [147,148]. The survivin gene contains a cyclin-dependent element, and derepression of this element allows the expression of survivin at transition phase G1/S; survivin levels remain high during mitosis since it participates in chromosome assembly at the mitotic spindle [149]. Survivin expression is regulated by the formation or dissociation of different Rb/E2F complexes throughout the cell cycle [150], and it is switched off by an increase in p53/p21 complexes [151]. Besides regulating mitosis progression, survivin has a

role in preventing apoptosis, possibly by impairing caspases activation and mitochondrial dysfunction [152].

Resveratrol decreased survivin levels by enhancing its degradation as well as reducing its transcription; this was associated with decreased proliferation and sensitization to chemotherapy [153]. Conversely, other authors assessed that resveratrol did not alter survivin or Bcl2 expression but it down-regulated other IAPs (cIAP1 and 2) related to apoptosis induction in a concentration-dependent manner [144].

4.5.3. Genomic regulation of cell cycle and apoptosis

NF- κ B is a transcription factor involved in titration of the balance between proliferation and apoptotic stress response. The RelA/NF- κ B family includes several proteins that, after release from cytosolic inhibitors (I- κ B) via phosphorylation, translocate to the nucleus. Dimeric transcriptional factors are activated by serine phosphorylation due to PKA, MAPKs or PKC ζ [154,155] and by acetylation due to different acetylases (including the CREB-binding protein, p300) [156]. Acetylation is a dynamic process that is reversed by deacetylases. Deacetylation reduces NF- κ B transcriptional potential and increases its affinity for the cytosolic inhibitor I- κ B [156]. The subset of activated genes depends on the combination of different monomers forming active NF- κ B as well as on combinatorial interactions with promoter-bound factors, according to stimuli and cell type [157].

AP1 is another transcriptional factor produced by a variety of dimeric combinations of proteins of the Jun and Fos families. AP1 is regulated via phosphorylation by MAPKs and interacts with other factors such as NF- κ B, CBP/p300 and Rb, thus regulating target genes common to NF- κ B. Like NF- κ B, AP1 is also considered to be a proliferation and tumor growth promoter [158].

Resveratrol was shown to inhibit NF- κ B activation; with some exceptions [159,160], this activation was associated with an antiproliferative action and with the induction of cell death [104,126,127]. NF- κ B controls the transcription of a variety of genes, including tumor-promoting COX2, iNOS, matrix metalloprotease (MMP9) and endothelial adhesion molecules [157]. The expression of these genes was reported to be down-regulated by resveratrol in different cell lines [60,85,86,98]. In addition, dietary administration of resveratrol in DMBA-induced tumor-bearing rats reduced tumor growth and decreased transcription of NF- κ B and of its regulated genes COX2 and MMP9 in tumor tissues [98]. Resveratrol was shown to inhibit NF- κ B, AP1 and their target genes regulating the activity of the upstream MAPKs. Resveratrol down-regulated TNF α induction of NF- κ B by blocking JNK and MEK activation [161], and it down-regulated phorbol myristate acetate (PMA)-induced NF- κ B by blocking JNK and PKC δ activation [60]. Similarly, resveratrol down-regulated AP1 induction by PMA and UV irradiation, possibly by upstream inhibition of Src and MAPKs [61].

Resveratrol inhibition of tyrosine kinases such as Raf [162] and Src [61] may be considered an upstream event that

opens access to multiple cascades. Studies either confirmed [61] or refuted [23] resveratrol inhibition of Src activity in vitro, questioning the possibility of a direct action.

The potential of resveratrol as an antiangiogenic molecule can be related to its inhibition of NF- κ B activation. In TNF α stimulated endothelial cells, resveratrol impaired NF- κ B activation only after prolonged and not acute treatment [163]. Moreover, low doses (1 μ M) of resveratrol blocked Src activation upon growth factor stimulation of human umbilical endothelial cells [23].

Another proposed mechanism for resveratrol inhibition of NF- κ B is via impairment of phosphorylation and activation of PKD. In vitro assays revealed that resveratrol inhibited, although weakly (IC₅₀ 100 μ M), PKD autophosphorylation, which is necessary for the kinase activation [164]. Protein kinase D is activated by Src- and Abl-mediated tyrosine phosphorylation and by PKC α serine phosphorylation; PKD activation is upstream to the NF- κ B survival response to oxidative stress [165]. Resveratrol inhibited PKC activity [61,166,167]. Some authors reported that resveratrol has a selective inhibitory effect on PKC α [60], and therefore, it can impair both PKD and downstream NF- κ B activation [168]. It is still unclear if resveratrol affects directly PKC δ activity, since the evidence of resveratrol competition for phorbol ester binding domain (IC₅₀, 2 μ M) [167] and kinase domain (IC₅₀, 90 μ M) [169] has been obtained studying, respectively, classic PKC isoforms or an isozymes mixture from rat brain.

Recently, it has been demonstrated that resveratrol is a powerful activator of sirtuins transcription and function [170]. Sirtuins are a nicotinamide adenosine dinucleotide (NAD)-dependent class of deacetylases responsible for regulating the response to DNA damage and gene silencing processes of aging and survival [171,172]. Resveratrol activated human sirtuin 1 (SIRT1) and this mediated RelA/p65 deacetylation, thus inhibiting TNF α -induced NF- κ B transcription and sensitizing cells to apoptosis [173].

This evidence contrasts with the observations that SIRT1 deacetylated and inactivated p53 [174] in a concerted and still not defined equilibrium with the action of histone deacetylase HDAC1 [121], thus providing a possible mechanism of blocking apoptosis and promoting cell survival [121,175]. In addition, resveratrol enhanced the expression p300 [54], an acetylase that activates NF- κ B [135,156] and p53 [135]. In this sense, Yeung et al. [173] showed that resveratrol failed to inhibit and actually increased TNF α induction of NF- κ B when SIRT1 was pharmacologically inhibited, suggesting that its activity may mediate resveratrol down-regulation of NF- κ B; conversely, the overexpression of SIRT1 reversed TNF α -induced and possibly p300-mediated activation of NF- κ B.

From the reported data, it is possible to speculate that resveratrol controls two key gene transcriptional regulators: p300 and SIRT1. It remains to be assessed whether this action could be a direct one by increasing enzyme activity, as has been shown in vitro for sirtuin [170]. Both p300 and

SIRT1 activate transcriptional factors that trigger opposite pathways of apoptosis and proliferation. p300 activates (by acetylation) whereas SIRT1 inhibits (by deacetylation) both the proliferative NF- κ B and the apoptotic p53. This hypothesis requires further investigation that may be important in elucidating a reported dual effect of resveratrol as an inducer of cell death as well as of proliferation (see following paragraph).

Egr1 is a transcriptional factor that activates genes related to cell growth and differentiation. Egr1 belongs to a group of early growth response genes (e.g., p53, Rb, growth factors, multidrug resistance MDR1, *c-jun*, *c-fos*). It is regulated by forming complexes with cofactors, such as CBP (p300) acetylases [136]. It is activated in response to cytokines, stress and cytotoxic agents and involved in proliferation and in cell death pathways [176,177]. Mitogen-activated protein kinases may be upstream mediators of Egr1 transcription [178,179].

Resveratrol was shown to activate Egr1. Egr1 can bind p21 promoter in vivo, and antisense Egr1 mRNA impaired resveratrol-induced p21 up-regulation [180]. Moreover, Egr1 transcription is mediated by ERKs, since specific inhibitors of these kinases block the resveratrol-induced increases in Egr1 [112,181]. It is interesting to note that resveratrol-induced early increase in the expression of Egr1 was not related to transcriptional events; conversely, a delayed and sustained expression of Egr1 resulted as a mediated event that was inhibited by cycloheximide [180]. This supports the hypothesis that Egr1 is a direct resveratrol target.

These observations suggest that cellular response to resveratrol involves gene transcription-mediated promotion of parallel or overlapping and potentiating cascades of events. By modulating upstream tyrosine and serine kinases, resveratrol may regulate the activation of transcriptional factors directed to clusters of genes responsible for inducing cell cycle arrest and eventually apoptosis. This hypothesis is summarized in Fig. 6.

4.5.4. Sphingolipid signaling

Ceramide is a sphingolipid mediator of intracellular signals, normally present in membranes in a complexed form as sphingomyelin or gangliosides. In conditions of stress and aging, there is an increased production of ceramide by de novo synthesis and release by hydrolysis of complex sphingolipids. Ceramide interacts, either directly or indirectly, with a variety of intracellular targets, leading to differentiation, cell cycle arrest and apoptosis.

Recently, resveratrol was shown to promote intracellular accumulation of ceramide in breast and prostate cancer cells [182,183]. Resveratrol enhanced the de novo synthesis of this sphingolipid by increasing the activity of the rate-limiting enzyme. Moreover, inhibitors of ceramide formation (enzyme blockers) rescued cells from resveratrol-induced apoptosis. This observation identified a new important checkpoint in the actions of resveratrol. Although this polyphenol triggers multiple pathways, ceramide production

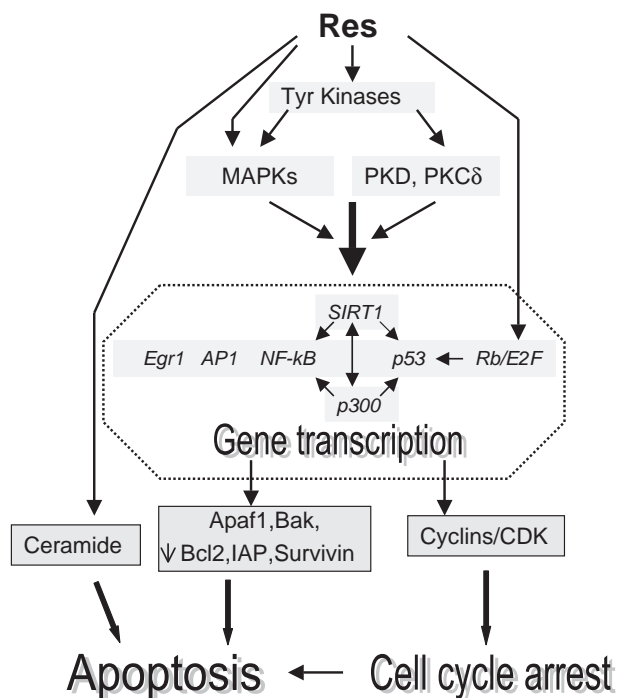


Fig. 6. Pathways in which resveratrol modulates cell cycle arrest and apoptosis.

may be a common step that drives cells toward irreversible death (Fig. 6).

4.5.5. Drug transporters

In addition to interfering with cell cycle control, resveratrol affects multiple targets that are involved directly or indirectly in apoptotic pathways and chemotherapy responsiveness. An intriguing hypothesis is that resveratrol, like other flavonoids, is able to overcome drug resistance of tumors that express multidrug resistance-associated proteins (MRP). These ATP-dependent pumps efflux chemotherapeutics out of cells [184].

Resveratrol was reported to interact with the breast cancer resistance protein (BCRP): it competed with other substrates for BCRP, impairing its activity and caused the intracellular accumulation of drugs [185]. It is important to note, however, that BCRP is also normally expressed in placenta in order to protect the fetus from maternal blood estrogens and at the luminal surface of the blood–brain barrier.

5. Effects of resveratrol on tumoral versus normal cells

A few reports have offered an interesting perspective on resveratrol actions on normal versus malignant cells. One study that compared different leukemia cell lines and bone marrow progenitor cells found that the IC_{50} of resveratrol for proliferation inhibition varied almost twofold: 34 μ M in leukemia and 59 μ M in hematopoietic cells. Moreover, in a colony formation assay performed after a pulse of resveratrol (80 μ M for 20 h), there was a significant difference in

the ability to proliferate between hematopoietic progenitors and leukemia cells. Human fibroblasts transformed with SV40 virus (WI38VA) were sensitive to resveratrol modulation of pro- versus antiapoptotic genes whereas normal fibroblasts were not [186].

Not only in leukemia but also in breast cancer, resveratrol induced apoptosis via CD95-dependent caspase activation (anti-CD95 antibody rescued from apoptosis). Both tumors constitutively expressed CD95, and resveratrol promoted CD95-ligand expression, which acted as an autocrine apoptosis inducer. Interestingly, peripheral blood lymphocytes, although CD95-positive, did not respond to resveratrol by ligand expression and therefore were insensitive to the compound cytotoxicity [130]. A slight difference in sensitivity to resveratrol has been reported between tongue squamous carcinoma cells and normal gingival fibroblasts [128], possibly related to the lower proliferation rate (therefore, higher resistance) of normal cells versus immortalized or highly proliferating tumoral cells.

These promising results contrast with those of other studies in which resveratrol exhibited similar effects on normal and neoplastic cells. Comparable cytostatic and cytotoxic potentials have been observed in breast cancer cell lines and in immortalized mammary epithelial cells [51].

6. Resveratrol double identity: prosurvival versus prodeath

What is particularly intriguing about resveratrol is its dual action. In plants, resveratrol is an antioxidant, protecting against damage induced by light exposure. However, its overproduction, triggered by excessive stress, can be considered as a hypersensitivity reaction that may lead to cell death. Considering the bioactivity of resveratrol in humans, dietary assumption may impair tumorigenesis by inhibiting the formation of toxic cellular metabolites and enhance the activity of sirtuins known to be related to cell survival. However, resveratrol is also able to arrest cell cycle and to induce apoptosis in many human tumor cell lines as well as in animals *in vivo*. What is the thread that connects these two signaling pathways that are commonly considered as diverging cascades?

Being a phytoestrogen, resveratrol activated steroid receptor-mediated proliferative pathways, as has been shown in osteoblasts [187] and breast cancer cells [48]. Nonetheless, antiproliferative effects have been shown to occur independently of ER expression or transcriptional activity [47].

NF- κ B and AP1 are activated in response to a variety of proliferative or stress signals [188]. Resveratrol inhibits NF- κ B activation and proliferative responses, as previously discussed. In addition, Manna et al. [59] reported that resveratrol inhibited NF- κ B activation in human myeloid lymphoma in response to apoptotic stimuli such as LPS and ceramide, therefore, impairing caspases activation and apoptosis. Resveratrol blocked p65 subunit phosphorylation

and nuclear translocation, thereby inhibiting NF- κ B-mediated activation of JNK and phosphorylation of MEK. It is to note that at least 4 h of preincubation with resveratrol is required for this effect [59].

Pretreatment with low doses (4–8 μ M) of resveratrol for 2–4 h prevented H₂O₂-induced apoptosis [59,189]. Reactive oxygen species, including superoxide O₂⁻ and H₂O₂, are maintained at nontoxic concentrations by intracellular antioxidant systems. If on one hand high concentrations of ROS create a necrotic stress, a slight increase may represent instead a proliferative stimulus. This often becomes an acquired survival and growth-promoting mechanisms in cancer cells [190]. It has been demonstrated that a moderate increase in O₂⁻ counteracts cell death and that the apoptotic potential of H₂O₂ is strengthened by its abilities to decrease the intracellular concentration of O₂⁻ and to reduce cytosolic pH [191,192]. Human leukemia cells, pretreated for 2 h with resveratrol at low doses, exhibited a significant increase in intracellular O₂⁻ that impaired H₂O₂-induced acidification, caspases activation and apoptosis [189].

In 2003, Howitz et al. [170] observed, in a yeast model, that resveratrol activated sirtuin (SIRT2, yeast homologue of human SIRT1). In yeast, sirtuins are activated in response to calorie restriction, which prolongs the cell life span. Resveratrol effects on sirtuins have also been studied in animal and human models. In HEK 293 cells, low doses of resveratrol (0.5 μ M) partially protected from radiation-induced apoptosis; this was associated with deacetylation and inactivation of p53 due to SIRT1 activation, although this protection effect was reversed when cells were treated at higher concentrations (50 μ M) [170]. Resveratrol directly regulated the activity in vitro and lowered the K_m of recombinant human SIRT1 for its acetylated substrate and for NAD⁺ [170]. In rat adipocytes, resveratrol (50–100 μ M) stimulated lipolysis and fatty acid mobilization from triglycerides, probably by binding DNA and preventing peroxisome proliferator-activated receptor (PPAR γ)-induced gene transcription [193]. Elongation of the life span in yeast and protection from cell death or white fat mobilization in mammalian cells represent responses to stress mediated by sirtuins. Resveratrol may mimic this stimulus and activate sirtuin-mediated intracellular events in order to counteract the stressing agent and allow the cell to live. On the other hand, dietary restriction inhibits signaling pathways (e.g., involving MAPKs and AP1) related to proliferation and cancer promotion [194,195]; similarly, resveratrol affects these targets in a variety of human tumors by acting as an antitumor agent.

These observations suggest that resveratrol has different effects according to cellular conditions, specific cell molecular settings and finally the concentration used. Resveratrol up-regulated the expression and/or enhanced the activity of transcriptional regulators: acetylase-transferases such as p300 and deacetylases such as sirtuin. Each of these two may exert opposite effects, as discussed previously. In this scenario, it is possible that in cells with altered metabolism

and unbalanced expression of proliferative over apoptotic regulators or subjected to stress factors (such as chemotherapeutics or cytokines), the equilibrium between the two opposite pathways results unbalanced thus particularly sensitive to resveratrol. Moreover, epigenetic regulation of gene transcription (DNA methylation and histone methylation, phosphorylation, acetylation and ubiquitination) is frequently associated with cancer promotion and progression [196], and the activity of deacetylases, by targeting transcriptional factors among other substrates, contributes significantly to cell cycle regulation [197]. Resveratrol modulation on acetylation/deacetylation mechanism requires further investigation to define applicable dose and tumor responsiveness and it might to turn out to be an upstream event that promotes the transcription of those sets of genes responsible for triggering signaling cascades.

7. Conclusions

In the last decade, the number of studies on resveratrol has dramatically increased from five publications in 1993 to over 200 in 2003. This is not surprising, as resveratrol is interesting for its participation in both prosurvival and prodeath cellular mechanisms, favoring the preservation of the functional status of cells and possibly elongating a cell life span, and inducing the death of cells whose physiological conditions have become deranged. Therefore, an increasing number of researchers are joining the challenge to unravel the mysteries of this fascinating and promising molecule, hopefully with success.

References

- [1] Langeake P, Pryce RJ. A new class of phytoalexins from grapevines. *Experientia* 1977;33:151–2.
- [2] Dixon RA. Natural products and plant disease resistance. *Nature* 2001;411:843–7.
- [3] Nakaune R, Hamamoto H, Imada J, Akutsu K, Hibi T. A novel ABC transporter gene, PMR5, is involved in multidrug resistance in the phytopathogenic fungus *Penicillium digitatum*. *Mol Genet Genomics* 2002;267:179–85.
- [4] Heath MC. Nonhost resistance and nonspecific plant defenses. *Curr Opin Plant Biol* 2000;3:315–9.
- [5] Regev-Shoshani G, Shoseyov O, Bilkis I, Kerem Z. Glycosylation of resveratrol protects it from enzymic oxidation. *Biochem J* 2003;374:157–63.
- [6] Sanders TH, McMichael Jr RW, Hendrix KW. Occurrence of resveratrol in edible peanuts. *J Agric Food Chem* 2000;48:1243–6.
- [7] Rimando AM, Kalt W, Magee JB, Dewey J, Ballington JR. Resveratrol, pterostilbene, and piceatannol in vaccinium berries. *J Agric Food Chem* 2004;52:4713–9.
- [8] Matsuda H, Tomohiro N, Hiraba K, Harima S, Ko S, Matsuo K, et al. Study on anti-Oketsu activity of rhubarb II. Anti-allergic effects of stilbene components from *Rheum undulati* Rhizoma (dried rhizome of *Rheum undulatum* cultivated in Korea). *Biol Pharm Bull* 2001;24:264–7.
- [9] Nonomura S, Kanagawa H, Makimoto A. Chemical constituents of polygonaceous plants. I. Studies on the components of Ko-J O-Kon. (*Polygonum cuspidatum* Sieb Et Zucc.). *Yakugaku Zasshi* 1963;83:988–90.

- [10] Hathway DE. The use of hydroxystilbene compounds as taxonomic tracers in the genus *Eucalyptus*. *Biochem J* 1962;83:80–4.
- [11] Oleszek W, Sitek M, Stochmal A, Piacente S, Pizza C, Cheeke P. Resveratrol and other phenolics from the bark of *Yucca schidigera roezl*. *J Agric Food Chem* 2001;49:747–52.
- [12] Likhitwitayawuid K, Sawasdee K, Kirtikara K. Flavonoids and stilbenoids with COX-1 and COX-2 inhibitory activity from *Dracaena loureiri*. *Planta Med* 2002;68:841–3.
- [13] Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997;275:218–20.
- [14] Surh YJ. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 2003;3:768–80.
- [15] Kelloff GJ, Crowell JA, Steele VE, Lubet RA, Malone WA, Boone CW, et al. Progress in cancer chemoprevention: development of diet-derived chemopreventive agents. *J Nutr* 2000;130:467S–71S.
- [16] Fremont L. Biological effects of resveratrol. *Life Sci* 2000;66:663–73.
- [17] Belguendouz L, Fremont L, Gozzelino MT. Interaction of trans-resveratrol with plasma lipoproteins. *Biochem Pharmacol* 1998;55:811–6.
- [18] Frankel EN, Kanner J, German JB, Parks E, Kinsella JE. Inhibition of oxidation of human low-density lipoprotein by phenolic substances in red wine. *Lancet* 1993;341:454–7.
- [19] Kerry NL, Abbey M. Red wine and fractionated phenolic compounds prepared from red wine inhibit low density lipoprotein oxidation in vitro. *Atherosclerosis* 1997;135:93–102.
- [20] Wang Z, Huang Y, Zou J, Cao K, Xu Y, Wu JM. Effects of red wine and wine polyphenol resveratrol on platelet aggregation in vivo and in vitro. *Int J Mol Med* 2002;9:77–9.
- [21] Rotondo S, Rajtar G, Manarini S, Celardo A, Rotillo D, de Gaetano G, et al. Effect of *trans*-resveratrol, a natural polyphenolic compound, on human polymorphonuclear leukocyte function. *Br J Pharmacol* 1998;123:1691–9.
- [22] Li HF, Chen SA, Wu SN. Evidence for the stimulatory effect of resveratrol on Ca(2+)-activated K⁺ current in vascular endothelial cells. *Cardiovasc Res* 2000;45:1035–45.
- [23] Lin MT, Yen ML, Lin CY, Kuo ML. Inhibition of vascular endothelial growth factor-induced angiogenesis by resveratrol through interruption of Src-dependent vascular endothelial cadherin tyrosine phosphorylation. *Mol Pharmacol* 2003;64:1029–36.
- [24] Pendurthi UR, Rao LV. Resveratrol suppresses agonist-induced monocyte adhesion to cultured human endothelial cells. *Thromb Res* 2002;106:243–8.
- [25] Aggarwal BB, Kumar A, Bharti AC. Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 2003;23:363–98.
- [26] Linden KG, Carpenter PM, McLaren CE, Barr RJ, Hite P, Sun JD, et al. Chemoprevention of nonmelanoma skin cancer: experience with a polyphenol from green tea. *Recent Results Cancer Res* 2003;163:165–71 [discussion 264–166].
- [27] Gusman J, Malonne H, Atassi G. A reappraisal of the potential chemopreventive and chemotherapeutic properties of resveratrol. *Carcinogenesis* 2001;22:1111–7.
- [28] Andlauer W, Kolb J, Siebert K, Furst P. Assessment of resveratrol bioavailability in the perfused small intestine of the rat. *Drugs Exp Clin Res* 2000;26:47–55.
- [29] Kuhnle G, Spencer JP, Chowrimootoo G, Schroeter H, Debnam ES, Srai SK, et al. Resveratrol is absorbed in the small intestine as resveratrol glucuronide. *Biochem Biophys Res Commun* 2000;272:212–7.
- [30] Bertelli A, Bertelli AA, Gozzini A, Giovannin L. Plasma and tissue resveratrol concentrations and pharmacological activity. *Drugs Exp Clin Res* 1998;24:133–8.
- [31] Soleas GJ, Angelini M, Grass L, Diamandis EP, Goldberg DM. Absorption of *trans*-resveratrol in rats. *Methods Enzymol* 2001;335:145–54.
- [32] Asensi M, Medina I, Ortega A, Carretero J, Bano MC, Obrador E, et al. Inhibition of cancer growth by resveratrol is related to its low bioavailability. *Free Radic Biol Med* 2002;33:387–98.
- [33] Marier JF, Vachon P, Gritsas A, Zhang J, Moreau JP, Ducharme MP. Metabolism and disposition of resveratrol in rats: extent of absorption, glucuronidation, and enterohepatic recirculation evidenced by a linked-rat model. *J Pharmacol Exp Ther* 2002;302:369–73.
- [34] Vitrac X, Desmouliere A, Brouillaud B, Krisa S, Deffieux G, Barthe N, et al. Distribution of [14C]-*trans*-resveratrol, a cancer chemopreventive polyphenol, in mouse tissues after oral administration. *Life Sci* 2003;72:2219–33.
- [35] Kaldas MI, Walle UK, Walle T. Resveratrol transport and metabolism by human intestinal Caco-2 cells. *J Pharm Pharmacol* 2003;55:307–12.
- [36] Lancon A, Delma D, Osman H, Thenot JP, Jannin B, Latruffe N. Human hepatic cell uptake of resveratrol: involvement of both passive diffusion and carrier-mediated process. *Biochem Biophys Res Commun* 2004;316:1132–7.
- [37] Goldberg DM, Yan J, Soleas GJ. Absorption of three wine-related polyphenols in three different matrices by healthy subjects. *Clin Biochem* 2003;36:79–87.
- [38] Krasnow MN, Murphy TM. Polyphenol glucosylating activity in cell suspensions of grape (*Vitis vinifera*). *J Agric Food Chem* 2004;52:3467–72.
- [39] Day AJ, DuPont MS, Ridley S, Rhodes M, Rhodes MJ, Morgan MR, et al. Deglycosylation of flavonoid and isoflavonoid glycosides by human small intestine and liver beta-glucosidase activity. *FEBS Lett* 1998;436:71–5.
- [40] Meng X, Maliakal P, Lu H, Lee MJ, Yang CS. Urinary and plasma levels of resveratrol and quercetin in humans, mice, and rats after ingestion of pure compounds and grape juice. *J Agric Food Chem* 2004;52:935–42.
- [41] Yu C, Shin YG, Kosmeder JW, Pezzuto JM, van Breemen RB. Liquid chromatography/tandem mass spectrometric determination of inhibition of human cytochrome P450 isozymes by resveratrol and resveratrol-3-sulfate. *Rapid Commun Mass Spectrom* 2003;17:307–13.
- [42] de Santi C, Pietrabissa A, Mosca F, Pacifici GM. Glucuronidation of resveratrol, a natural product present in grape and wine, in the human liver. *Xenobiotica* 2000;30:1047–54.
- [43] Manzocco L, Calligaris S, Nicoli MC. Assessment of pro-oxidant activity of foods by kinetic analysis of crocin bleaching. *J Agric Food Chem* 2002;50:2767–71.
- [44] Jordan VC. Antiestrogens and selective estrogen receptor modulators as multifunctional medicines. 2. Clinical considerations and new agents. *J Med Chem* 2003;46:1081–111.
- [45] Davis SR, Dalais FS, Simpson ER, Murkies AL. Phytoestrogens in health and disease. *Recent Prog Horm Res* 1999;54:185–210 [discussion 210–181].
- [46] Ganry O. Phytoestrogen and breast cancer prevention. *Eur J Cancer Prev* 2002;11:519–22.
- [47] Levenson AS, Gehm BD, Pearce ST, Horiguchi J, Simons LA, Ward III JE, et al. Resveratrol acts as an estrogen receptor (ER) agonist in breast cancer cells stably transfected with ER alpha. *Int J Cancer* 2003;104:587–96.
- [48] Gehm BD, McAndrews JM, Chien PY, Jameson JL. Resveratrol, a polyphenolic compound found in grapes and wine, is an agonist for the estrogen receptor. *Proc Natl Acad Sci U S A* 1997;94:14138–43.
- [49] Gehm BD, Levenson AS, Liu H, Lee EJ, Amundsen BM, Cushman M, et al. Estrogenic effects of resveratrol in breast cancer cells expressing mutant and wild-type estrogen receptors: role of AF-1 and AF-2. *J Steroid Biochem Mol Biol* 2004;88:223–34.
- [50] Bhat KPL, Kosmeder II JW, Pezzuto JM. Biological effects of resveratrol. *Antioxid Redox Signal* 2001;3:1041–64.
- [51] Mgbonyebi OP, Russo J, Russo IH. Antiproliferative effect of synthetic resveratrol on human breast epithelial cells. *Int J Oncol* 1998;12:865–9.

- [52] Lu R, Serrero G. Resveratrol, a natural product derived from grape, exhibits antiestrogenic activity and inhibits the growth of human breast cancer cells. *J Cell Physiol* 1999;179:297–304.
- [53] Bhat KP, Pezzuto JM. Resveratrol exhibits cytostatic and antiestrogenic properties with human endometrial adenocarcinoma (Ishikawa) cells. *Cancer Res* 2001;61:6137–44.
- [54] Narayanan BA, Narayanan NK, Stoner GD, Bullock BP. Interactive gene expression pattern in prostate cancer cells exposed to phenolic antioxidants. *Life Sci* 2002;70:1821–39.
- [55] Mitchell SH, Zhu W, Young CY. Resveratrol inhibits the expression and function of the androgen receptor in LNCaP prostate cancer cells. *Cancer Res* 1999;59:5892–5.
- [56] Kuwajewala N, Cifuentes E, Gautam S, Menon M, Barrack ER, Reddy GP. Resveratrol induces prostate cancer cell entry into S phase and inhibits DNA synthesis. *Cancer Res* 2002;62:2488–92.
- [57] Brownson DM, Azios NG, Fuqua BK, Dharmawardhane SF, Mabry TJ. Flavonoid effects relevant to cancer. *J Nutr* 2002;132:3482S–9S.
- [58] Kousteni S, Han L, Chen JR, Almeida M, Plotkin LI, Bellido T, et al. Kinase-mediated regulation of common transcription factors accounts for the bone-protective effects of sex steroids. *J Clin Invest* 2003;111:1651–64.
- [59] Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF-kappa B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. *J Immunol* 2000;164:6509–19.
- [60] Woo JH, Lim JH, Kim YH, Suh SI, Min do S, Chang JS, et al. Resveratrol inhibits phorbol myristate acetate-induced matrix metalloproteinase-9 expression by inhibiting JNK and PKC delta signal transduction. *Oncogene* 2004;23:1845–53.
- [61] Yu R, Hebbar V, Kim DW, Mandelkar S, Pezzuto JM, Kong AN. Resveratrol inhibits phorbol ester and UV-induced activator protein 1 activation by interfering with mitogen-activated protein kinase pathways. *Mol Pharmacol* 2001;60:217–24.
- [62] Zhang S, Cao HJ, Davis FB, Tang HY, Davis PJ, Lin HY. Oestrogen inhibits resveratrol-induced post-translational modification of p53 and apoptosis in breast cancer cells. *Br J Cancer* 2004;91:178–85.
- [63] Coleman KM, Smith CL. Intracellular signaling pathways: non-genomic actions of estrogens and ligand-independent activation of estrogen receptors. *Front Biosci* 2001;6:D1379–91.
- [64] Damianaki A, Bakogeorgou E, Kampa M, Notas G, Hatzoglou A, Panagiotou S, et al. Potent inhibitory action of red wine polyphenols on human breast cancer cells. *J Cell Biochem* 2000;78:429–41.
- [65] El-Mowafy AM, Alkhalaf M. Resveratrol activates adenylyl-cyclase in human breast cancer cells: a novel, estrogen receptor-independent cytostatic mechanism. *Carcinogenesis* 2003;24:869–73.
- [66] Donato MT, Castell JV. Strategies and molecular probes to investigate the role of cytochrome P450 in drug metabolism: focus on in vitro studies. *Clin Pharmacokinet* 2003;42:153–78.
- [67] Marchand A, Barouki R, Garlatti M. Regulation of NAD(P)H:quinone oxidoreductase 1 gene expression by CYP1A1 activity. *Mol Pharmacol* 2004;65:1029–37.
- [68] Mikstacka R, Gnojowski J, Baer-Dubowska W. Effect of natural phenols on the catalytic activity of cytochrome P450 2E1. *Acta Biochim Pol* 2002;49:917–25.
- [69] Ciolino HP, Yeh GC. Inhibition of aryl hydrocarbon-induced cytochrome P-450 1A1 enzyme activity and CYP1A1 expression by resveratrol. *Mol Pharmacol* 1999;56:760–7.
- [70] Shou M, Korzekwa KR, Krausz KW, Buters JT, Grogan J, Goldfarb I, et al. Specificity of cDNA-expressed human and rodent cytochrome P450s in the oxidative metabolism of the potent carcinogen 7,12-dimethylbenz[a]anthracene. *Mol Carcinog* 1996;17:241–9.
- [71] Chang TK, Chen J, Lee WB. Differential inhibition and inactivation of human CYP1 enzymes by *trans*-resveratrol: evidence for mechanism-based inactivation of CYP1A2. *J Pharmacol Exp Ther* 2001;299:874–82.
- [72] Szaefer H, Cichocki M, Brauze D, Baer-Dubowska W. Alteration in phase I and II enzyme activities and polycyclic aromatic hydrocarbons — DNA adduct formation by plant phenolics in mouse epidermis. *Nutr Cancer* 2004;48:70–7.
- [73] Aktan F. iNOS-mediated nitric oxide production and its regulation. *Life Sci* 2004;75:639–53.
- [74] Balligand JL, Cannon PJ. Nitric oxide synthases and cardiac muscle. Autocrine and paracrine influences. *Arterioscler Thromb Vasc Biol* 1997;17:1846–58.
- [75] Ignarro LJ. Endothelium-derived nitric oxide: actions and properties. *FASEB J* 1989;3:31–6.
- [76] Kubes P, Suzuki M, Granger DN. Nitric oxide: an endogenous modulator of leukocyte adhesion. *Proc Natl Acad Sci U S A* 1991;88:4651–5.
- [77] Sarkar R, Meinberg EG, Stanley JC, Gordon D, Webb RC. Nitric oxide reversibly inhibits the migration of cultured vascular smooth muscle cells. *Circ Res* 1996;78:225–30.
- [78] Kroncke KD, Fehsel K, Kolb-Bachofen V. Inducible nitric oxide synthase in human diseases. *Clin Exp Immunol* 1998;113:147–56.
- [79] Vieira H, Kroemer G. Mitochondria as targets of apoptosis regulation by nitric oxide. *IUBMB Life* 2003;55:613–6.
- [80] Radisavljevic Z. Inactivated tumor suppressor Rb by nitric oxide promotes mitosis in human breast cancer cells. *J Cell Biochem* 2004;92:1–5.
- [81] Ohshima H. Genetic and epigenetic damage induced by reactive nitrogen species: Implications in carcinogenesis. *Toxicol Lett* 2003;140–141:99–104.
- [82] Xia YF, Liu LP, Zhong CP, Geng JG. NF-kappaB activation for constitutive expression of VCAM-1 and ICAM-1 on B lymphocytes and plasma cells. *Biochem Biophys Res Commun* 2001;289:851–6.
- [83] Roman V, Billard C, Kern C, Ferry-Dumazet H, Izard JC, Mohammad R, et al. Analysis of resveratrol-induced apoptosis in human B-cell chronic leukaemia. *Br J Haematol* 2002;117:842–51.
- [84] Orallo F, Alvarez E, Camina M, Leiro JM, Gomez E, Fernandez P. The possible implication of *trans*-resveratrol in the cardioprotective effects of long-term moderate wine consumption. *Mol Pharmacol* 2002;61:294–302.
- [85] Hsieh TC, Juan G, Darzynkiewicz Z, Wu JM. Resveratrol increases nitric oxide synthase, induces accumulation of p53 and p21(WAF1/CIP1), and suppresses cultured bovine pulmonary artery endothelial cell proliferation by perturbing progression through S and G2. *Cancer Res* 1999;59:2596–601.
- [86] Tsai SH, Lin-Shiau SY, Lin JK. Suppression of nitric oxide synthase and the down-regulation of the activation of NFkappaB in macrophages by resveratrol. *Br J Pharmacol* 1999;126:673–80.
- [87] Wang D, Dubois RN. Cyclooxygenase-2: a potential target in breast cancer. *Semin Oncol* 2004;31:64–73.
- [88] Rouzer CA, Mamett LJ. Mechanism of free radical oxygenation of polyunsaturated fatty acids by cyclooxygenases. *Chem Rev* 2003;103:2239–304.
- [89] Szwczuk LM, Forti L, Stivala LA, Penning TM. Resveratrol is a peroxidase-mediated inactivator of COX-1 but not COX-2: a mechanistic approach to the design of COX-1 selective agents. *J Biol Chem* 2004;279:22727–37.
- [90] Vane JR. Inhibition of prostaglandin synthesis as a mechanism of action for aspirin-like drugs. *Nat New Biol* 1971;231:232–5.
- [91] Wu KK. Aspirin and other cyclooxygenase inhibitors: New therapeutic insights. *Semin Vasc Med* 2003;3:107–12.
- [92] Gasparini G, Longo R, Sarmiento R, Morabito A. Inhibitors of cyclo-oxygenase 2: A new class of anticancer agents? *Lancet Oncol* 2003;4:605–15.
- [93] Tsujii M, DuBois RN. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell* 1995;83:493–501.
- [94] Tsujii M, Kawano S, DuBois RN. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. *Proc Natl Acad Sci U S A* 1997;94:3336–40.

- [95] Subbaramaiah K, Chung WJ, Michaluart P, Telang N, Tanabe T, Inoue H, et al. Resveratrol inhibits cyclooxygenase-2 transcription and activity in phorbol ester-treated human mammary epithelial cells. *J Biol Chem* 1998;273:21875–82.
- [96] McAdam BF, Catella-Lawson F, Mardini IA, Kapoor S, Lawson JA, FitzGerald GA. Systemic biosynthesis of prostacyclin by cyclooxygenase (COX)-2: the human pharmacology of a selective inhibitor of COX-2. *Proc Natl Acad Sci U S A* 1999;96:272–7.
- [97] O'Leary KA, de Pascual-Tereasa S, Needs PW, Bao YP, O'Brien NM, Williamson G. Effect of flavonoids and vitamin E on cyclooxygenase-2 (COX-2) transcription. *Mutat Res* 2004;551:245–54.
- [98] Banerjee S, Bueso-Ramos C, Aggarwal BB. Suppression of 7,12-dimethylbenz(a)anthracene-induced mammary carcinogenesis in rats by resveratrol: role of nuclear factor-kappaB, cyclooxygenase 2, and matrix metalloprotease 9. *Cancer Res* 2002;62:4945–54.
- [99] Murray AW. Recycling the cell cycle: cyclins revisited. *Cell* 2004;116:221–34.
- [100] Ahmad N, Adhami VM, Afaq F, Feyes DK, Mukhtar H. Resveratrol causes WAF-1/p21-mediated G(1)-phase arrest of cell cycle and induction of apoptosis in human epidermoid carcinoma A431 cells. *Clin Cancer Res* 2001;7:1466–73.
- [101] Ragione FD, Cucciolla V, Borriello A, Pietra VD, Racioppi L, Soldati G, et al. Resveratrol arrests the cell division cycle at S/G2 phase transition. *Biochem Biophys Res Commun* 1998;250:53–8.
- [102] Haider UG, Sorescu D, Griendling KK, Vollmar AM, Dirsch VM. Resveratrol increases serine15-phosphorylated but transcriptionally impaired p53 and induces a reversible DNA replication block in serum-activated vascular smooth muscle cells. *Mol Pharmacol* 2003;63:925–32.
- [103] Mnjoyan ZH, Fujise K. Profound negative regulatory effects by resveratrol on vascular smooth muscle cells: a role of p53-p21(WAF1/CIP1) pathway. *Biochem Biophys Res Commun* 2003;311:546–52.
- [104] Estrov Z, Shishodia S, Faderl S, Harris D, Van Q, Kantarjian HM, et al. Resveratrol blocks interleukin-1beta-induced activation of the nuclear transcription factor NF-kappaB, inhibits proliferation, causes S-phase arrest, and induces apoptosis of acute myeloid leukemia cells. *Blood* 2003;102:987–95.
- [105] Latruffe N, Delmas D, Jannin B, Malki MC, Passilly-Degrace P, Berlot JP. Molecular analysis on the chemopreventive properties of resveratrol, a plant polyphenol microcomponent. *Int J Mol Med* 2002;10:755–60.
- [106] Pozo-Guisado E, Alvarez-Barrientos A, Mulero-Navarro S, Santiago-Josefat B, Fernandez-Salguero PM. The antiproliferative activity of resveratrol results in apoptosis in MCF-7 but not in MDA-MB-231 human breast cancer cells: cell-specific alteration of the cell cycle. *Biochem Pharmacol* 2002;64:1375–86.
- [107] Joe AK, Liu H, Suzui M, Vural ME, Xiao D, Weinstein IB. Resveratrol induces growth inhibition, S-phase arrest, apoptosis, and changes in biomarker expression in several human cancer cell lines. *Clin Cancer Res* 2002;8:893–903.
- [108] Larrosa M, Tomas-Barberan FA, Espin JC. Grape polyphenol resveratrol and the related molecule 4-hydroxystilbene induce growth inhibition, apoptosis, S-phase arrest, and upregulation of cyclins A, E, and B1 in human SK-Mel-28 melanoma cells. *J Agric Food Chem* 2003;51:4576–84.
- [109] Fulda S, Debatin KM. Sensitization for anticancer drug-induced apoptosis by the chemopreventive agent resveratrol. *Oncogene* 2004;23:6702–11.
- [110] Liang YC, Tsai SH, Chen L, Lin-Shiau SY, Lin JK. Resveratrol-induced G2 arrest through the inhibition of CDK7 and p34CDC2 kinases in colon carcinoma HT29 cells. *Biochem Pharmacol* 2003;65:1053–60.
- [111] Carbo N, Costelli P, Baccino FM, Lopez-Soriano FJ, Argiles JM. Resveratrol, a natural product present in wine, decreases tumour growth in a rat tumour model. *Biochem Biophys Res Commun* 1999;254:739–43.
- [112] Della Ragione F, Cucciolla V, Criniti V, Indaco S, Borriello A, Zappia V. Antioxidants induce different phenotypes by a distinct modulation of signal transduction. *FEBS Lett* 2002;532:289–94.
- [113] Park JW, Choi YJ, Jang MA, Lee YS, Jun DY, Suh SI, et al. Chemopreventive agent resveratrol, a natural product derived from grapes, reversibly inhibits progression through S and G2 phases of the cell cycle in U937 cells. *Cancer Lett* 2001;163:43–9.
- [114] Bruder JL, Hsieh T, Lerea KM, Olson SC, Wu JM. Induced cytoskeletal changes in bovine pulmonary artery endothelial cells by resveratrol and the accompanying modified responses to arterial shear stress. *BMC Cell Biol* 2001;2:1.
- [115] Nielsen M, Ruch RJ, Vang O. Resveratrol reverses tumor-promoter-induced inhibition of gap-junctional intercellular communication. *Biochem Biophys Res Commun* 2000;275:804–9.
- [116] Pucci B, Kasten M, Giordano A. Cell cycle and apoptosis. *Neoplasia* 2000;2:291–9.
- [117] Lin HY, Shih A, Davis FB, Tang HY, Martino LJ, Bennett JA, et al. Resveratrol induced serine phosphorylation of p53 causes apoptosis in a mutant p53 prostate cancer cell line. *J Urol* 2002;168:748–55.
- [118] Adhami VM, Afaq F, Ahmad N. Involvement of the retinoblastoma (pRb)-E2F/DP pathway during antiproliferative effects of resveratrol in human epidermoid carcinoma (A431) cells. *Biochem Biophys Res Commun* 2001;288:579–85.
- [119] Appella E, Anderson CW. Post-translational modifications and activation of p53 by genotoxic stresses. *Eur J Biochem* 2001;268:2764–72.
- [120] Goodman RH, Smolik S. CBP/p300 in cell growth, transformation, and development. *Genes Dev* 2000;14:1553–77.
- [121] Gu W, Luo J, Brooks CL, Nikolaev AY, Li M. Dynamics of the p53 acetylation pathway. *Novartis Found Symp* 2004;259:197–205 [discussion 205–197, 223–195].
- [122] Ashcroft M, Taya Y, Vousden KH. Stress signals utilize multiple pathways to stabilize p53. *Mol Cell Biol* 2000;20:3224–33.
- [123] Andreassen PR, Lohez OD, Lacroix FB, Margolis RL. Tetraploid state induces p53-dependent arrest of nontransformed mammalian cells in G1. *Mol Biol Cell* 2001;12:1315–28.
- [124] Oren M. Regulation of the p53 tumor suppressor protein. *J Biol Chem* 1999;274:36031–4.
- [125] Haupt S, Berger M, Goldberg Z, Haupt Y. Apoptosis — the p53 network. *J Cell Sci* 2003;116:4077–85.
- [126] Kim YA, Lee WH, Choi TH, Rhee SH, Park KY, Choi YH. Involvement of p21WAF1/CIP1, pRB, Bax and NF-kappaB in induction of growth arrest and apoptosis by resveratrol in human lung carcinoma A549 cells. *Int J Oncol* 2003;23:1143–9.
- [127] Narayanan BA, Narayanan NK, Re GG, Nixon DW. Differential expression of genes induced by resveratrol in LNCaP cells: P53-mediated molecular targets. *Int J Cancer* 2003;104:204–12.
- [128] Babich H, Reisbaum AG, Zuckerbraun HL. In vitro response of human gingival epithelial S-G cells to resveratrol. *Toxicol Lett* 2000;114:143–53.
- [129] Gautam SC, Xu YX, Dumaguin M, Janakiraman N, Chapman RA. Resveratrol selectively inhibits leukemia cells: a prospective agent for ex vivo bone marrow purging. *Bone Marrow Transplant* 2000;25:639–45.
- [130] Clement MV, Hirpara JL, Chawdhury SH, Pervaiz S. Chemopreventive agent resveratrol, a natural product derived from grapes, triggers CD95 signaling-dependent apoptosis in human tumor cells. *Blood* 1998;92:996–1002.
- [131] She QB, Bode AM, Ma WY, Chen NY, Dong Z. Resveratrol-induced activation of p53 and apoptosis is mediated by extracellular-signal-regulated protein kinases and p38 kinase. *Cancer Res* 2001;61:1604–10.
- [132] Chang F, Steelman LS, Shelton JG, Lee JT, Navolanic PM, Blalock WL, et al. Regulation of cell cycle progression and apoptosis by the Ras/Raf/MEK/ERK pathway (Review). *Int J Oncol* 2003;22:469–80.

- [133] Niles RM, McFarland M, Weimer MB, Redkar A, Fu YM, Meadows GG. Resveratrol is a potent inducer of apoptosis in human melanoma cells. *Cancer Lett* 2003;190:157–63.
- [134] Gayther SA, Batley SJ, Linger L, Bannister A, Thorpe K, Chin SF, et al. Mutations truncating the EP300 acetylase in human cancers. *Nat Genet* 2000;24:300–3.
- [135] Culmsee C, Siewe J, Junker V, Retiounskaia M, Schwarz S, Camandola S, et al. Reciprocal inhibition of p53 and nuclear factor-kappaB transcriptional activities determines cell survival or death in neurons. *J Neurosci* 2003;23:8586–95.
- [136] Silverman ES, Du J, Williams AJ, Wadgaonkar R, Drazen JM, Collins T. cAMP-response-element-binding-protein-binding protein (CBP) and p300 are transcriptional co-activators of early growth response factor-1 (Egr-1). *Biochem J* 1998;336(Pt 1):183–9.
- [137] Harbour JW, Dean DC. Rb function in cell-cycle regulation and apoptosis. *Nat Cell Biol* 2000;2:E65–7.
- [138] Ginsberg D. E2F1 pathways to apoptosis. *FEBS Lett* 2002;529:122–5.
- [139] Fontecave M, Lepoivre M, Elleingand E, Gerez C, Guittet O. Resveratrol, a remarkable inhibitor of ribonucleotide reductase. *FEBS Lett* 1998;421:277–9.
- [140] Sun NJ, Woo SH, Cassady JM, Snapka RM. DNA polymerase and topoisomerase II inhibitors from *Psoralea corylifolia*. *J Nat Prod* 1998;61:362–6.
- [141] Cho DI, Koo NY, Chung WJ, Kim TS, Ryu SY, Im SY, et al. Effects of resveratrol-related hydroxystilbenes on the nitric oxide production in macrophage cells: structural requirements and mechanism of action. *Life Sci* 2002;71:2071–82.
- [142] Fesik SW. Insights into programmed cell death through structural biology. *Cell* 2000;103:273–82.
- [143] Tinhofer I, Bernhard D, Senfter M, Anether G, Loeffler M, Kroemer G, et al. Resveratrol, a tumor-suppressive compound from grapes, induces apoptosis via a novel mitochondrial pathway controlled by Bcl-2. *FASEB J* 2001;15:1613–5.
- [144] Park JW, Choi YJ, Suh SI, Baek WK, Suh MH, Jin IN, et al. Bcl-2 overexpression attenuates resveratrol-induced apoptosis in U937 cells by inhibition of caspase-3 activity. *Carcinogenesis* 2001;22:1633–9.
- [145] Pellicchia M, Reed JC. Inhibition of antiapoptotic Bcl-2 family proteins by natural polyphenols: new avenues for cancer chemoprevention and chemotherapy. *Curr Pharm Des* 2004;10:1387–98.
- [146] Delmas D, Rebe C, Lacour S, Filomenko R, Athias A, Gambert P, et al. Resveratrol-induced apoptosis is associated with Fas redistribution in the rafts and the formation of a death-inducing signaling complex in colon cancer cells. *J Biol Chem* 2003;278:41482–90.
- [147] Altieri DC. Survivin, versatile modulation of cell division and apoptosis in cancer. *Oncogene* 2003;22:8581–9.
- [148] Deveraux QL, Reed JC. IAP family proteins — suppressors of apoptosis. *Genes Dev* 1999;13:239–52.
- [149] Lens SM, Wolthuis RM, Klompmaker R, Kaur J, Agami R, Brummelkamp T, et al. Survivin is required for a sustained spindle checkpoint arrest in response to lack of tension. *EMBO J* 2003;22:2934–47.
- [150] Jiang Y, Saavedra HL, Holloway MP, Leone G, Altura RA. Aberrant regulation of survivin by the RB/E2F family of proteins. *J Biol Chem* 2004;279:40511–20.
- [151] Hoffman WH, Biade S, Zilfou JT, Chen J, Murphy M. Transcriptional repression of the antiapoptotic survivin gene by wild type p53. *J Biol Chem* 2002;277:3247–57.
- [152] Castedo M, Perfettini JL, Roumier T, Andreau K, Medema R, Kroemer G. Cell death by mitotic catastrophe: a molecular definition. *Oncogene* 2004;23:2825–37.
- [153] Hayashibara T, Yamada Y, Nakayama S, Harasawa H, Tsuruda K, Sugahara K, et al. Resveratrol induces downregulation in survivin expression and apoptosis in HTLV-1-infected cell lines: a prospective agent for adult T cell leukemia chemotherapy. *Nutr Cancer* 2002;44:193–201.
- [154] Duran A, Diaz-Meco MT, Moscat J. Essential role of RelA Ser311 phosphorylation by zetaPKC in NF-kappaB transcriptional activation. *EMBO J* 2003;22:3910–8.
- [155] Vermeulen L, De Wilde G, Van Damme P, Vanden Berghe W, Haegeman G. Transcriptional activation of the NF-kappaB p65 subunit by mitogen- and stress-activated protein kinase-1 (MSK1). *EMBO J* 2003;22:1313–24.
- [156] Zhong H, May MJ, Jimi E, Ghosh S. The phosphorylation status of nuclear NF-kappa B determines its association with CBP/p300 or HDAC-1. *Mol Cell* 2002;9:625–36.
- [157] Chen LF, Greene WC. Shaping the nuclear action of NF-kappaB. *Nat Rev Mol Cell Biol* 2004;5:392–401.
- [158] Chinenov Y, Kerppola TK. Close encounters of many kinds: fos–Jun interactions that mediate transcription regulatory specificity. *Oncogene* 2001;20:2438–52.
- [159] Shen F, Chen SJ, Dong XJ, Zhong H, Ti LY, Cheng GF. Suppression of IL-8 gene transcription by resveratrol in phorbol ester treated human monocytic cells. *J Asian Nat Prod Res* 2003;5:151–7.
- [160] Jang JH, Surh YJ. Protective effects of resveratrol on hydrogen peroxide-induced apoptosis in rat pheochromocytoma (PC12) cells. *Mutat Res* 2001;496:181–90.
- [161] Manna SK, Mukhopadhyay A, Aggarwal BB. IFN-alpha suppresses activation of nuclear transcription factors NF-kappa B and activator protein 1 and potentiates TNF-induced apoptosis. *J Immunol* 2000;165:4927–34.
- [162] Gao S, Liu GZ, Wang Z. Modulation of androgen receptor-dependent transcription by resveratrol and genistein in prostate cancer cells. *Prostate* 2004;59:214–25.
- [163] Pellegatta F, Bertelli AA, Staels B, Duhem C, Fulgenzi A, Ferrero ME. Different short- and long-term effects of resveratrol on nuclear factor-kappaB phosphorylation and nuclear appearance in human endothelial cells. *Am J Clin Nutr* 2003;77:1220–8.
- [164] Stewart JR, Christman KL, O'Brian CA. Effects of resveratrol on the autophosphorylation of phorbol ester-responsive protein kinases: inhibition of protein kinase D but not protein kinase C isozyme autophosphorylation. *Biochem Pharmacol* 2000;60:1355–9.
- [165] Storz P, Doppler H, Toker A. Protein kinase Cdelta selectively regulates protein kinase D-dependent activation of NF-kappaB in oxidative stress signaling. *Mol Cell Biol* 2004;24:2614–26.
- [166] Nixon JS, Bishop J, Bradshaw D, Davis PD, Hill CH, Elliott LH, et al. The design and biological properties of potent and selective inhibitors of protein kinase C. *Biochem Soc Trans* 1992;20:419–25.
- [167] Slater SJ, Seiz JL, Cook AC, Stagliano BA, Buzas CJ. Inhibition of protein kinase C by resveratrol. *Biochim Biophys Acta* 2003;1637:59–69.
- [168] Storz P, Doeppler H, Toker A. Activation loop phosphorylation controls protein kinase D-dependent activation of nuclear factor kappa-B. *Mol Pharmacol* 2004;66:870–9.
- [169] Stewart JR, Ward NE, Ioannides CG, O'Brian CA. Resveratrol preferentially inhibits protein kinase C-catalyzed phosphorylation of a cofactor-independent, arginine-rich protein substrate by a novel mechanism. *Biochemistry* 1999;38:13244–51.
- [170] Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, et al. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 2003;425:191–6.
- [171] Guarente L. Sir2 links chromatin silencing, metabolism, and aging. *Genes Dev* 2000;14:1021–6.
- [172] Cohen HY, Miller C, Bitterman KJ, Wall NR, Hekking B, Kessler B, et al. Caloric restriction promotes mammalian cell survival by inducing the SIRT1 deacetylase. *Science* 2004;305:390–2.
- [173] Yeung F, Hoberg JE, Ramsey CS, Keller MD, Jones DR, Frye RA, et al. Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 2004;23:2369–80.
- [174] Vaziri H, Dessain SK, Ng Eaton E, Imai SI, Frye RA, Pandita TK, et al. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell* 2001;107:149–59.

- [175] Luo J, Nikolaev AY, Imai S, Chen D, Su F, Shiloh A, et al. Negative control of p53 by Sir2alpha promotes cell survival under stress. *Cell* 2001;107:137–48.
- [176] Carleton M, Haks MC, Smeele SA, Jones A, Belkowski SM, Berger MA, et al. Early growth response transcription factors are required for development of CD4(-)CD8(-) thymocytes to the CD4(+)CD8(+) stage. *J Immunol* 2002;168:1649–58.
- [177] Thiel G, Cibelli G. Regulation of life and death by the zinc finger transcription factor Egr-1. *J Cell Physiol* 2002;193:287–92.
- [178] Schwachtgen JL, Houston P, Campbell C, Sukhatme V, Braddock M. Fluid shear stress activation of egr-1 transcription in cultured human endothelial and epithelial cells is mediated via the extracellular signal-related kinase 1/2 mitogen-activated protein kinase pathway. *J Clin Invest* 1998;101:2540–9.
- [179] Granet C, Boutahar N, Vico L, Alexandre C, Lafage-Proust MH. MAPK and SRC-kinases control EGR-1 and NF-kappa B inductions by changes in mechanical environment in osteoblasts. *Biochem Biophys Res Commun* 2001;284:622–31.
- [180] Ragione FD, Cucciolla V, Criniti V, Indaco S, Borriello A, Zappia V. p21Cip1 gene expression is modulated by Egr1: a novel regulatory mechanism involved in the resveratrol antiproliferative effect. *J Biol Chem* 2003;278:23360–8.
- [181] Quinones A, Dobberstein KU, Rainov NG. The egr-1 gene is induced by DNA-damaging agents and non-genotoxic drugs in both normal and neoplastic human cells. *Life Sci* 2003;72:2975–92.
- [182] Scarlatti F, Sala G, Somenzi G, Signorelli P, Sacchi N, Ghidoni R. Resveratrol induces growth inhibition and apoptosis in metastatic breast cancer cells via de novo ceramide signaling. *FASEB J* 2003;17:2339–41.
- [183] Sala G, Minutolo F, Macchia M, Sacchi N, Ghidoni R. Resveratrol structure and ceramide-associated growth inhibition in prostate cancer cells. *Drugs Exp Clin Res* 2003;29:263–9.
- [184] Wortelboer HM, Usta M, van der Velde AE, Boersma MG, Spenkelink B, van Zanden JJ, et al. Interplay between MRP inhibition and metabolism of MRP inhibitors: the case of curcumin. *Chem Res Toxicol* 2003;16:1642–51.
- [185] Cooray HC, Janvilisri T, van Veen HW, Hladky SB, Barrand MA. Interaction of the breast cancer resistance protein with plant polyphenols. *Biochem Biophys Res Commun* 2004;317:269–75.
- [186] Lu J, Ho CH, Ghai G, Chen KY. Resveratrol analog, 3,4,5,4'-tetrahydroxystilbene, differentially induces proapoptotic p53/Bax gene expression and inhibits the growth of transformed cells but not their normal counterparts. *Carcinogenesis* 2001;22:321–8.
- [187] Mizutani K, Ikeda K, Kawai Y, Yamori Y. Resveratrol stimulates the proliferation and differentiation of osteoblastic MC3T3-E1 cells. *Biochem Biophys Res Commun* 1998;253:859–63.
- [188] Young MR, Yang HS, Colburn NH. Promising molecular targets for cancer prevention: AP-1, NF-kappa B and Pcd4. *Trends Mol Med* 2003;9:36–41.
- [189] Ahmad KA, Clement MV, Pervaiz S. Pro-oxidant activity of low doses of resveratrol inhibits hydrogen peroxide-induced apoptosis. *Ann N Y Acad Sci* 2003;1010:365–73.
- [190] Burdon RH. Superoxide and hydrogen peroxide in relation to mammalian cell proliferation. *Free Radic Biol Med* 1995;18:775–94.
- [191] Clement MV, Pervaiz S. Reactive oxygen intermediates regulate cellular response to apoptotic stimuli: an hypothesis. *Free Radic Res* 1999;30:247–52.
- [192] Pervaiz S, Ramalingam JK, Hirpara JL, Clement MV. Superoxide anion inhibits drug-induced tumor cell death. *FEBS Lett* 1999;459:343–8.
- [193] Picard F, Kurtev M, Chung N, Topark-Ngarm A, Senawong T, Machado De Oliveira R, et al. Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 2004;429:771–6.
- [194] Liu Y, Wang W, Hawley J, Birt DF. Adrenalectomy abrogates reduction of 12-O-tetradecanoylphorbol-13-acetate-induced extracellular signal-regulated protein kinase activity in the epidermis of dietary energy-restricted SENCAR mice: implications of glucocorticoid hormone. *Cancer Epidemiol Biomarkers Prev* 2002;11:299–304.
- [195] Przybyszewski J, Yaktine AL, Duysen E, Blackwood D, Wang W, Au A, et al. Inhibition of phorbol ester-induced AP-1-DNA binding, c-Jun protein and *c-jun* mRNA by dietary energy restriction is reversed by adrenalectomy in SENCAR mouse epidermis. *Carcinogenesis* 2001;22:1421–7.
- [196] Kopelovich L, Crowell JA, Fay JR. The epigenome as a target for cancer chemoprevention. *J Natl Cancer Inst* 2003;95:1747–57.
- [197] Johnstone RW. Histone-deacetylase inhibitors: novel drugs for the treatment of cancer. *Nat Rev Drug Discov* 2002;1:287–99.